theranos	LDT Validation Report	Theranos Lipase Assay	Rev:
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Description	Validation Report for Modified Siemens Assay of Lipase in Lithium Heparin Plasma		
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Validation of Modified Siemens Lipase Assay

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Lipase Plasma Assay

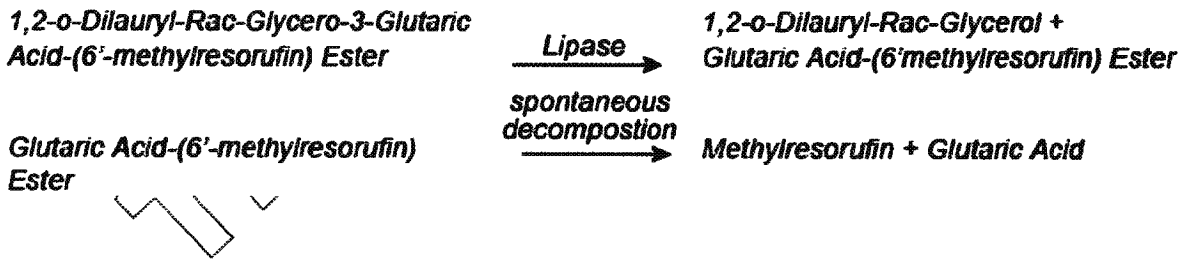
Overview

Lipases are enzymes that hydrolyze glycerol esters of long-chain fatty acids and produce fatty acids and 2-acylglycerol. Bile salts and a cofactor, colipase, are required for full catalytic activity and greatest specificity. The pancreas is the primary source of serum lipase. Both lipase and colipase are synthesized in the pancreatic acinar cells and secreted by the pancreas in roughly equimolar amounts. Lipase is filtered and reabsorbed by the kidneys. Pancreatic injury results in increased serum lipase levels.

I. Method Principle

The chromogenic lipase substrate, DGGMR, (1,2-o-dilauryl-rac-glycero-3-glutaric acid-(6'-methylresorufin)ester) is cleaved by the catalytic action of lipase to form 1,2-o-dilauryl-rac-glycerol and an unstable intermediate, glutaric acid-(6'-methylresorufin)ester. This decomposes spontaneously in an alkaline solution to form glutaric acid and methylresorufin. The lipase activity in the specimen is proportional to the production of methylresorufin in the reaction and is determined spectrophotometrically.

Reaction Equation



II. Definitions and Abbreviations

The following definitions and abbreviations are used in this document and related documents and attachments:

a. **Accuracy:** Accuracy is defined by CLSI as the closeness of agreement between a test result and an accepted reference value. Method accuracy is used in a different sense by the American Association of Pharmaceutical Scientists where it is expressed as percent relative error (%RE). Trueness, a related CLSI term, is the closeness of agreement between the average of a number of replicate measured quantity values and a reference quantity value.

b. **Analyte:** Component represented in the name of a measurable quantity. The closely related term measureand is defined as the particular quantity subject to measurement.

c. **Analytical sensitivity:** There are several alternative uses of this term. Most commonly, and for the purposes of this Validation Plan, it is used interchangeably with limit of detection. It is also used to describe the ability of an analytical method to assess small variations of the concentration of an analyte, such as the slope of the calibration curve (IUPAC).

d. **Analytical specificity:** Ability of a test or procedure to correctly identify or quantify an entity, including in the presence of interfering substance(s) or phenomena.

e. **Calibration:** Set of operations that establish, under specified conditions, the relationship between values of quantities indicated by a measuring instrument or measuring system, or values represented by a material measure or a reference material, and the corresponding values realized by standards. Under CLIA, calibration refers to the process of testing and adjusting an instrument, kit, or test system, to provide a known relationship between the measurement response and the value of the substance being measured by the test procedure (42 CFR 493.1217).

f. **Calibrator:** A substance, material, or article intended to be used to establish the measurement relationships of a diagnostic medical device.

g. **CLIA:** Clinical Laboratory Improvement Amendments of 1988. Congressional legislation that defined and requires specific quality assurance practices in clinical laboratories.

h. **CLSI:** Clinical and Laboratory Standards Institute.

i. **Coefficient of Variation:** The ratio of the standard deviation to the average, often multiplied by 100 and expressed as a percentage, abbreviated as %CV .

j. **Colorimetry:** A technique used to determine the concentration of colored compound(s) in solution.

k. **Interfering substance:** A substance or quantity thereof that is not the measurand but that affects the result of the measurement.

l. **IUPAC:** International Union of Pure and Applied Chemistry

m. **LDT:** Laboratory –developed Test.

n. **Linearity:** Linearity is the ability of a quantitative analytical method to provide results that are directly proportional to the concentrations of an analyte in test samples, within a given measuring interval. It is an important parameter to confirm when evaluating an analytical method because it verifies correct interpolation of results between points.

o. **LMR:** Lower end of the measuring range is the lowest level at which defined conditions, including all stated characteristic of the method, are met.

p. **LoB:** Limit of Blank is the highest value in a series of results on a sample that contains no analyte.

q. **LoD:** Limit of Detection is the lowest amount of analyte in a sample that can be detected with stated probability, although perhaps not quantified as an exact value.

r. **LoQ:** When used without a prefix, the Limit of Quantitation is the lowest actual concentration at which an analyte is reliably detected and at which uncertainty of the test result is less than or equal to the goal set by the manufacturer or laboratory. The term may also be used with prefixes L for lower (LLOQ) and U for upper (ULOQ), respectively. Note: $LoB < LoD \leq LoQ$.

s. **Matrix:** All components of a material system, except the analyte. A specimen matrix is the biological milieu in which an analyte exists (e.g., plasma, serum, urine, or other body fluids).

t. **Measuring Interval (reportable range; analytical measurement range or AMR):** A measuring interval consists of all numeric values between the lower and upper numeric values for which a method can produce quantitative results suitable for clinical use. Where applicable, a linearity study is frequently used to establish or verify the measuring interval that can be reported for a measurement method. Alternatively, the lower limit of the measuring interval may be assigned as the LoQ (LLOQ).

u. **Precision:** Precision is the closeness of agreement between indications or measured quantity values obtained by replicate measurements on the same or similar objects under specified conditions. It is usually expressed numerically in terms of standard deviation (SD) or percent Coefficient of Variation (%CV).

v. **Reference interval:** The interval between and including two reference limits. It is common practice to define a reference limit so a stated fraction of the reference values is less than or equal, or greater than or equal, to the respective upper or lower limit.

w. **SOP:** Standard Operating Procedure.

x. **Spectrophotometry:** The quantitative measurement of the transmission (or reflection) properties of a material as a function of wavelength.

y. **Testing System:** The entirety of the testing process, including instrument, sample, reagents, supplies, and procedures. Personnel are sometimes included in the definition.

III. Pre-clinical Validation

a. Analytical Measurement Range

i. Limits of Blank, Detection and Quantitation

The limits of blank, detection, and quantitation were determined to be 5.0 U/L, 8.4 U/L and 11.7 U/L respectively.

Limit of blank

CLSI guideline EP17-A section 4.3.1

Level	Number of samples	N	Mean	SD
Blank	1	20	1.8	2.0
Alpha	5%			
Parametric LoB	5.0			

Limit of detection

CLSI guideline EP17-A section 4.3.2

Level	Number of samples	N	Pooled SD
Low	1	20	2.1
Beta	5%		
Parametric LoD	8.4		

Limit of quantitation

CLSI guideline EP17-A section 5.1

Level	Number of samples	N
Low	1	20
Bias	1.7	
Pooled imprecision	2.1	
95% total error	5.7	
Allowable error	2	

The lower limit of quantitation has been established at 11.7 U/L (17.6% CV and 117.0% recovery).

ii. Linearity

The Analytical Measurement Range (AMR) including linear measurement interval has been determined by Siemens. Refer to the Analytical Range section of the manufacturer product information insert for additional details.

b. Analytical Specificity

c. The analytical specificity for this assay was determined by testing the effect of hemoglobin (100 mg/dL), bilirubin (10 mg/dL) and triglycerides (400 mg/dL) on plasma samples spiked with the interferents and then compared with un-spiked controls. Lipase concentration at which the interference testing was performed at was 87.5 U/L. Non-interference was defined as the mean result from testing of spiked samples within 10% of the mean of the un-spiked samples. Recoveries were within 97.7% to 100.7% (see table below).

Table 1. Interference Testing For Lipase.

Analyte (mg/dL)	% Recovery		
	Interferent		
	Bilirubin (10 mg/dL)	Hemoglobin (100 mg/dL)	Triglycerides (400 mg/dL)
Lipase	97.7	98.2	100.7

No significant interference was observed

d. Precision

Level = Level 1

Number of observations	80
Number of runs	40
Number of days	20
Runs per day	2
Replicates per run	2
Mean	19.4

	SD	95% CI	CV	Allowable Total SD
Repeatability	0.0	0.0 to 0.0	0.0%	-
Between-run	3.3		17.1%	-
Between-day	1.7		8.7%	-
Within-laboratory	3.7	3.0 to 4.8	19.1%	5.8

Imprecision is less than allowable total imprecision: 30% upto 100U/L then 20%.

Level = Level 2

Number of observations	80
Number of runs	40
Number of days	20
Runs per day	2
Replicates per run	2
Mean	51.9

	SD	95% CI	CV	Allowable Total SD
Repeatability	0.0	0.0 to 0.0	0.0%	-
Between-run	4.0		7.6%	-
Between-day	2.0		3.8%	-
Within-laboratory	4.4	3.6 to 5.7	8.6%	15.6

Imprecision is less than allowable total imprecision: 30% upto 100U/L then 20%.

Level = Level 3

Number of observations	80
Number of runs	40
Number of days	20
Runs per day	2
Replicates per run	2
Mean	181.6

	SD	95% CI	CV	Allowable Total SD
Repeatability	0.0	0.0 to 0.0	0.0%	-
Between-run	4.0		2.2%	-
Between-day	3.1		1.7%	-
Within-laboratory	5.1	4.1 to 6.7	2.8%	36.3

Imprecision is less than allowable total imprecision: 30% upto 100U/L then 20%.

The percent CV reported as zeros in the above precision summary are most likely a consequence of rounding the values in StatisPro.

IV. Clinical Validation

a. Method Comparison with Predicate (Accuracy/Comparability)

To test the accuracy of the assay on the Theranos System, forty eight (48) unique patient samples were screened on the predicate method (Siemens, Advia) and on the Theranos method. One sample was excluded as an outlier (mean absolute difference greater than 4). Using the predicate method twenty two (22) values were within the reference range (6 - 51 U/L), none (0) value was below the reference range, and twenty five values (25) were above the reference range. Based on the results of the data examination, either a simple linear regression or alternative procedures were used to estimate expected (average) bias and the confidence interval of expected bias at the desired medical decision level(s) as per CLSI guidance EP09-A2. StatisPro was used for bias calculations. These estimates were compared with internal criteria to judge the acceptability of the Theranos method. Each sample was run in duplicate on the predicate, and the average used for comparison to the Theranos method. Some samples were stored before analysis on both methods. If the confidence interval for

the predicted bias includes the defined acceptable bias or if the acceptable bias is greater than the higher limit of the confidence interval of the predicted bias, then the data do not show that the bias of the Theranos method is different from the acceptable bias or there is a high probability (97%) that the predicated bias is acceptable, respectively. The acceptable bias at each medical decision level was determined based on the total allowable error (TEa) minus the measured precision at the level closest to that decision level. Total allowable error (TEa) was taken from American Proficiency Institute (API) peer proficiency testing criteria or CLIA proficiency testing criteria for acceptable analytical performance, as printed in the Federal Register February 28, 1992;57(40):7002-186, when available. The TEa for Lipase is 30%. The table below shows the allowable bias and precision at 2 levels (values shown in parentheses) and the corresponding closest medical decision limits.

Table 2. Allowable Bias and Precision at the Medical Decision Levels

Medical Decision Levels (U/L)	**50 (52)**	**210 (182)**
Precision (%)	**3.8**	**1.7**
Allowable Bias (%)	**26.2**	**28.3**

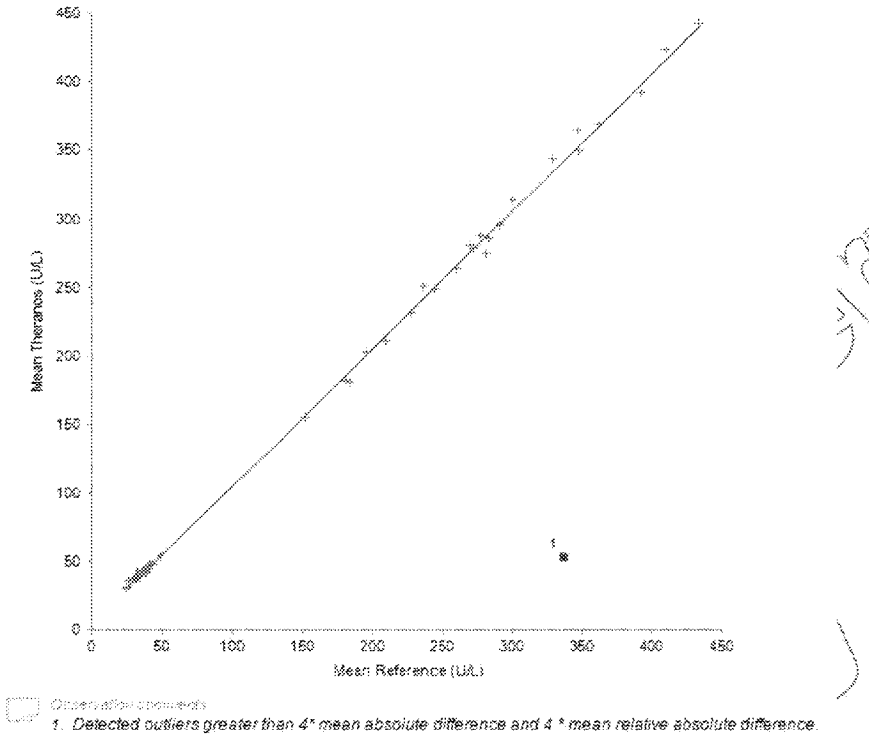


Figure 1. Graph showing Theranos method versus Predicate Method (Siemens Advia).

Simple linear regression was used to establish a slope, intercept and an r^2. The slope, intercept and clinical correlation were determined to be 1.00, 4.54 and 0.99 respectively.

Comparability

CLSI guideline EP09-A (Diff section) ?

Level ID	Value	Difference	SE	95% CI	Allowable difference
	50.000000	4.7148052	0.82842899	3.0462636 to 6.3833466	13.1000000
	200.000000	5.2255256	0.66121990	3.8937604 to 6.5572909	52.4000000
	500.000000	6.2469665	1.72570069	2.7712265 to 9.7227065	150.0000000

Difference is less than allowable bias: 26.2% upto 210U/L then 30%.

The difference between the two methods is not greater than the allowable difference. The performance requirement is verified.

b. Transference and Verification of Reference Interval (Venous)

Reference ranges were modified by applying the regression equation to the lower and upper reference limits of existing reference interval to generate a new reference range. New reference ranges were verified using a minimum of twenty (20) new normal subjects

New reference ranges were verified using a total of forty four (44) new normal subjects with matched Lithium heparin venous and finger sticks samples. For a reference range to pass verification, 95% of values should fall within the upper and lower reference limits and 5% or fewer values fall outside of the upper and lower reference limits. For venous verification 42 (95.5%) values fell within the new reference range and 2 (4.5%) values fell outside the new reference range. See graphs below for venous samples verification.

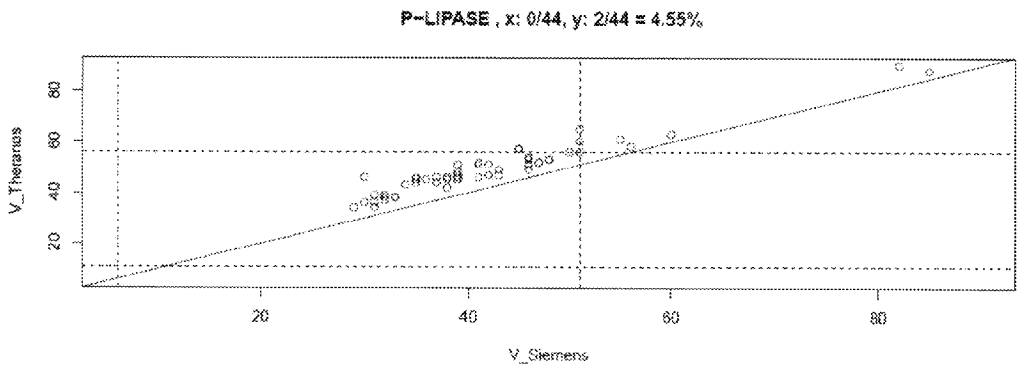


Figure 2. Graph showing venous sample reference range verification.

c. Verification of Reference Interval with Finger Stick Samples

New reference ranges were also verified with venous matched finger sticks (Lithium heparin) samples from a total of sixty (60) new normal subjects. The finger stick samples were collected in a Theranos blood collection device (BCD) configured with separate Lithium heparin and EDTA vessels. For finger stick verification 57 values (95.0 %) fell within the new reference range and 3 value (5.0 %) fell outside the new reference range. See graphs below for finger stick samples verification.

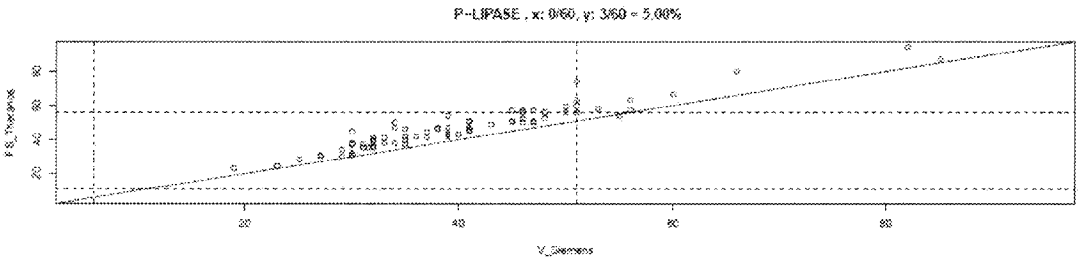


Figure 3. Graph showing Finger stick sample reference range verification.

The new reference range for finger stick lipase was determined to be 11 – 56 U/L.

VI. Stability

a. Reagents

On-board Reagent Stability

System	Stability
ADVIA 1200	12 days
ADVIA 1650/1800	10 days
ADVIA 2400	30 days

For all systems, unopened reagents are stable until the expiration date printed on the product label when stored at 2°C - 8°C. Do not freeze the reagents.

For additional details, refer to the Methods Introduction section of the system-specific Operator's Guide.

b. Sample

Plasma samples for lipase analysis are stable for 1 week at 2-8 °C, or at least 90 days at -20 °C.

c. Calibrators

Siemens Special Chemistry Calibrators should be stored at 2-8 °C, protected from light, and are stable until the expiration date on the vial label. Opened reconstituted calibrators are stable for 7 days, except for acid phosphatase, which is stable for at least 2 days.

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REVISION HISTORY			
Revision Level	**Effective Date**	**Initiator**	**ECO Number**
A	11/10/2013	A. Rosendorff	CL ECO-00118
Section Number	**Description and Justification of Changes**		
All	Initial Release		