

# Heterophile Antibodies IgM Assay Development Report

**Theranos, Inc.**

Aug. 29<sup>th</sup>, 2012

Prepared by: Salina Abusali

This development report contains Theranos Confidential Information and is being provided under the parties' Mutual Confidentiality Agreement. Any further dissemination, use or disclosure of the Report, in whole or in part, is strictly prohibited.

[ PAGE \\* MERGEFORMAT ]

## **Table of Contents**

[ TOC \o "1-3" \h \z \u ]

THP FM Internal Only

[ PAGE \\* MERGEFORMAT ]

[ TOC \h \z \c "Table" ]

THP FM Internal Only

[ PAGE \\* MERGEFORMAT ]

## **1. ASSAY INFORMATION[ TC "ASSAY INFORMATION" \f C \l "2" ]**

### **1.1 Assay Specifications[ TC "Assay Specifications" \f C \l "3" ]**

Heterophile antibodies are produced by the human immune system in response to EBV infection (infectious mononucleosis).

This assay is designed to qualitatively determine Heterophile antibodies (IgM) in human plasma and serum.

#### **1.1.1 Reference Assays [ TC "Reference Assays and Standards" \f C \l "3" ]**

The following commercial kits have been used in-house as predicate methods:

- Genzyme OSOM Mono Test (Cat# GZM-145)
- Status Mono (Life Sign LLC) (Cat# 84M30)

(Qualitative Detection of Infectious Mononucleosis Heterophile Antibodies in Whole Blood, Serum or Plasma)

#### **1.1.2 Materials and Methods[ TC "Materials and Methods" \f C \l "1" ]**

Heterophile antigen coated surface serves as the capture surface for the Anti-Heterophile antibody assay. The sample (plasma or serum) is diluted and then incubated on the capture surface for 10 minutes, the surface is washed, and then an alkaline phosphatase (AP)-labeled anti-human IgM antibody is incubated on the surface for 10 minutes. After the detection antibody incubation, another washing cycle is performed and the alkaline phosphatase substrate is incubated on the surface for 10 minutes, and the resulting chemiluminescence is read in Relative Light Units (RLU).

**Table [ SEQ Table \\* ARABIC ]: Materials**

Name	Supplier	Catalog #
Carbonate-Bicarbonate buffer	Sigma	C3041
Alkaline Phosphatase Labeling Kit	Dojindo	LK13-10
StabilZyme AP	Surmodics	SA01-1000
Blocking Buffer (3% BSA in TBS, 0.05% Sodium Azide)	Sigma (BSA, Fraction V, 99% Pure)	A3059-500G
In house Substrate	Theranos	
StatusFirst Mono Controls	Life Sign	200014

**Table [ SEQ Table \\* ARABIC ]: Antigens**

Antigen #	Vendor	Product Code	Description
1	East Coast Bio	L501L	Heterophile antigen
2	Fitzgerald	30-AH71	Mononucleosis protein antigen

**Table [ SEQ Table \\* ARABIC ]: Detection Antibodies**

DAb #	Supplier	Catalog #	Description
1	AbD Serotec	5278-5159	Mouse Anti Human IgM
2	Novus	NB7436	Human IgM Antibody
3	Novus	NB500-468	Human IgM, Fc Fragment Antibody (CH2)

## 2 ASSAY DEVELOPMENT[ TC "ASSAY OPTIMIZATION" \F C \L "2" ]

### 2.1 Capture Surface: Antigen Screen (MTP)

Clinical samples were screened on the Genzyme and Status mono kits to determine positive and negative Heterophile Antibodies IgM samples. Commercially available antigens were then screened with these positive and negative samples on a microtiter plate (MTP). Each antigen was prepared in Carbonate-Bicarbonate buffer for direct coating on the Nunc-384 well plates. All antigens were coated at 10 ug/ml. A sample dilution of 1:25 was added to the surface followed by wash steps. The detection antibody (D#1, 2, 3) concentration was 100 ng/ml in 3% BSA in TBS blocking buffer. Both Antigens #1 and #2 showed good modulation between positive and negative samples but antigen # 1 had a better modulation and hence Antigen #1 was chosen for further development. Direct coating of raw antigen had much better modulation than the ultraavidin coating of the biotinylated antigen because the biotinylation of the antigen (with SH group) destroyed the antigen structure and hence there was no modulation with positive and negative control with biotinylated antigen. The data is summarized in Table 4 and 5

**Table [ SEQ Table \\* ARABIC ]: Antigen Coating Methods (MTP)**

C#1 at 10ug/ml and D# 1 at 100ng/ml <b>Sample ID</b>	<b>Avidin Surface</b>		<b>Direct Coat</b>	
	<b>MeanValue</b>	<b>CV%</b>	<b>MeanValue</b>	<b>CV%</b>
Negative Control	7035	5	12073	10
Positive Control	2905	21	1045635	2.8
Negative Sample	575091	5	83432	9
Positive Sample	423953	6	1130514	6
No Sample*	737	9	726	19
<i>Positive control/negative control</i>	0		87	
<i>Positive control/Mean normal</i>	0		13	
<i>Mean positive /Mean normal</i>	1		14	

\*No Sample: Blocking buffer blank with detection antibody and substrate.

Table [ SEQ Table \\* ARABIC ]: Capture Surface Screen (MTP)

Antigen #	1		2	
[Antigen], ug/ml	10ug/ml		10ug/ml	
Sample#	Mean Value	CV%	Mean Value	CV%
Negative: #1	75442	9	172559	3
#2	91421	9	243502	9
<b>Mean Negative</b>	<b>83432</b>		<b>208031</b>	
Positive: #3	915185	11	982858	11
#4	1345842	1	1410887	5
<b>Mean Positive</b>	<b>1130514</b>		<b>1196873</b>	
Negative Control	12073	10	26772	5
Positive Control	1045635	2.8	1182337	11
No Sample*	726	19		
<b>Positive control/negative control</b>	87		44	
<b>Positive control/ Mean normal</b>	13		6	
<b>Mean positive /Mean normal</b>	14		6	

\*No Sample: Blocking buffer blank with detection antibody and substrate.

## 2.2 Capture Antigen Surface Titration on Theranos System[ TC " Capture Antigen Surface Antigen " \f C \l "1" ]

The direct coat antigen surface was titrated at levels: 10, 5 and 1 µg/mL. Table 6 summarizes the results of Antigen #1 and Detection Antibody # 1 at 100ng/mL. 1 µg/mL provides the best modulation between the pooled positive and pooled normal clinical samples and was finalized as the capture antigen surface concentration.

**Table [ SEQ Table \\* ARABIC ]: Capture Antigen Surface Titration**

	10 µg/mL		5 µg/mL		1 µg/mL	
Sample ID	Inter-Cartridge RLU					
	Mean	CV%	Mean	CV%	Mean	CV%
Positive Control	214917	18	190571	23	166526	18
Pooled Positive	2288791	9	2175271	12	2024979	8
Pooled normals	87461	17	66967	21	39811	14
Negative Control	2329	20	1889	9	1062	14
Positive control/negative control	92		101		157	
Positive control/ Mean normal	2		3		4	
Mean positive /Mean normal	26		32		51	

## 2.3 Effect of different Detection Antibodies on Theranos System

Antigen #1 was tested with the different detection antibodies. These antigens were screened on the Theranos system at 1 ug/ml direct coat. Three different detection antibody was being tested and the concentration was 100 ng/ml in 3% BSA Blocking buffer. Clinical samples were tested on the above mentioned commercial kits, and then used as the test set on the Theranos system. A sample dilution of 1:25 is manually done and the protocol was run at the 10, 10,10 minutes incubation time.

There was a significant improvement in terms of modulation between the three detection antibodies. Dab # 3 gave the best modulation with low background and was selected for further optimization. Dab # 1 is a possible back-up detection antibody, the data is summarized in Table 7

**Table [ SEQ Table \\* ARABIC ]: Detection Antibodies on Theranos System**

	Dab #1		Dab # 2		Dab # 3	
Sample ID	Inter-Cartridge RLU					
	Mean	CV%	Mean	CV%	Mean	CV%
Positive Control	213192	17	1595161	18	56309	19
Pooled Positive	1929907	13	3490887	7	1061400	25
Pooled normals	26710	18	286941	12	7247	15
Negative Control	1658	11	12410	26	763	21
Positive control/negative control	129		129		74	
Positive control/ Mean normal	8		6		8	
Mean positive /Mean normal	72		12		146	

## 2.4 Effect of Assay Diluent

Three commercially available blockers (SuperBlock®, StartingBlock™ and SeaBlock) and one in-house blocking buffer were tested as diluents for the assay. Data was compared to the control diluent which was the blocking buffer consisted of 3% BSA and 0.05% sodium azide in TBS. There was a not a lot of difference in modulation between each diluent and hence In house 3% BSA in TBS was chosen for further optimization. The data is summarized in Table 8

**Table [ SEQ Table \\* ARABIC ]: Effect of Assay Diluent**

Sample Type	Sample #	In House BB		Super Block		Starting Block		Sea Block	
		Mean RLU	CV%	Mean RLU	CV%	Mean RLU	CV%	Mean RLU	CV%
Negative Samples	1	3535	15	4476	18	3481	24	6900	23
	2	5489	13	5026	18	4227	23	6187	18
	3	2489	11	3928	19	4210	22	5255	17
	4	57796	5	37458	20	42791	12	28411	14
	5	1442	17	2084	15	1845	9	2730	16
	6	1959	9	9889	20	10037	13	11722	13
	7	539	15	2557	14	2284	18	3381	18
	Mean Negative	<b>10464</b>		<b>9345</b>		<b>9839</b>		<b>9226</b>	
Positive Samples	8	258955	24	139239	20	124808	28	153539	26
	9	817865	25	608016	6	573463	16	556823	21
	10	1631917	31	1579826	8	1714087	3	1821697	5
	11	1536862	21	1279139	17	1493682	1	1468091	19
	Mean Positive	<b>1061400</b>		<b>901555</b>		<b>976510</b>		<b>1000037</b>	
	Modulation	<b>101</b>		<b>96</b>		<b>99</b>		<b>108</b>	

## 2.5 HAMA and Rf Positive Sample Testing

5 HAMA positive and 5 Rf positive sera obtained from a commercial source were tested on the Theranos Heterophile Antibody IgM Assay and on the two commercial kits. Out of the 10 samples tested, all were negative for Heterophile antibodies IgM showing that our assay has excellent correlation. Table is summarized in Table 9

Table [ SEQ Table \\* ARABIC ]: HAMA and Rf positive sample screen

Samples	Inter-Cartridge		Theranos	Osom	Status
	Mean	CV%	Assay	Mono	Mono
<b>HAMA positive</b>				Test	
H2	9669	19	NEG	NEG	NEG
H4	7211	11	NEG	NEG	NEG
H5	6460	13	NEG	NEG	NEG
H6	10041	22	NEG	NEG	NEG
H7	15238	13	NEG	NEG	NEG
			NEG	NEG	NEG
<b>RF Positive</b>			NEG	NEG	NEG
RF 8	8062	25	NEG	NEG	NEG
RF 9	2780	14	NEG	NEG	NEG
RF 10	3941	13	NEG	NEG	NEG
RF 11	3788	9	NEG	NEG	NEG
RF 12	5665	12	NEG	NEG	NEG
<b>Mean</b>					
<b>Negative+2SD</b>	<b>52328</b>				

## 2.6 Specificity (Cross Reactivity Sample Testing)

Literature mentioned that Heterophile antibody assay could have cross reactivity with other infectious diseases like rubella, toxoplasma, HBsAg, Hep C, HIV1 and 2. Positive sera or QC controls of various infectious diseases were tested on the Theranos Heterophile Antibody IgM Assay and on the two commercial kits. All Samples came out negative showing that the Heterophile antibody assay has high specificity.

**Table [ SEQ Table \\* ARABIC ]: Cross reactivity testing with various infectious disease positive samples**

Samples	Inter-Cartridge		Theranos	Osom	Status
	Mean	CV%	Assay	Mono Test	Mono
Virotrol Mumz	1378	19	NEG	NEG	NEG
Hep B Antibody	5183	17	NEG	NEG	NEG
Anti HSV-1	2128	16	NEG	NEG	NEG
Total Anti HBC	3652	12	NEG	NEG	NEG
HBsAg	4643	5	NEG	NEG	NEG
Hep C Antibody	756	19	NEG	NEG	NEG
Anti Rubella	2061	9	NEG	NEG	NEG
Anti Toxoplasma	3434	20	NEG	NEG	NEG
Anti CMV	2989	19	NEG	NEG	NEG
HIV 1	4136	23	NEG	NEG	NEG
HIV 2	4067	15	NEG	NEG	NEG
<b>Mean Negative+2SD</b>	<b>52328</b>				

## 2.7 Effect of Detection Conjugate Stabilizer

Two commercial and one in house formulated alkaline phosphatase stabilizers were tested as detection antibody diluents, with the anti-human IgM DAb at 100 ng/mL. The samples were diluted 1:25 into 3% BSA in TBS Blocking Buffer. Signal modulation was best with StabilZyme. Table 11 summarizes the results of running the tests on 20 negative clinical samples and 5 positive clinical samples.

**Table [ SEQ Table \\* ARABIC ]: Effect of Detection Conjugate Stabilizer**

Sample Type	Theranos AP Conjugate Stabilizer		BioStab	StabilZyme
Negative Samples	Mean RLU	CV%	Mean RLU	CV %
			CV %	CV %

[ PAGE \\* MERGEFORMAT ]

1	2842	14	5577	20	1559	12
2	5503	20	9237	15	2425	13
3	3005	11	5538	23	1764	17
4	60102	6	78745	7	29791	9
5	4859	9	5783	19	3625	18
6	23567	17	31106	14	11325	10
7	12764	13	22162	17	4895	11
8	13314	18	17406	14	6448	6
9	3597	16	6705	8	2067	13
10	14781	9	18039	16	7255	15
11	11776	6	6039	19	2343	4
12	7599	12	10479	11	4252	6
13	3107	19	6647	9	2374	12
14	43749	9	50904	11	18927	5
15	1648	9	3620	20	952	15
16	4539	17	6519	13	2088	19
17	2239	14	4489	14	1098	11
18	21360	11	30756	9	8845	8
19	9370	12	14549	10	3582	17
20	5661	4	11364	12	2195	18
<b>Mean Negative</b>	<b>12769</b>		<b>17283</b>		<b>5890</b>	
<b>Positive Samples</b>	Mean RLU	CV%	Mean RLU	CV %	Mean RLU	CV %
1	273650	10	326918	7	123846	11
2	371538	7	529704	8	208698	2
3	871938	10	1019256	3	519362	3
4	1997198	18	2233970	5	1655136	5
5	1722123	8	1805897	13	1232998	14
<b>Mean Positive</b>	<b>1047289.267</b>		<b>1183149.01</b>		<b>748008</b>	
<b>Mean Negative +2 SD</b>	<b>42765</b>		<b>54793</b>		<b>20096</b>	
<b>Modulation (Mean Pos/ Mean Neg+2SD)</b>	<b>24</b>		<b>22</b>		<b>37</b>	

## 2.8 Detection antibody Titration

The AP conjugated detection antibody was titrated in StabilZyme. The best modulation between the positive and negative control was achieved with 25 ng/mL of the anti-IgM Dab. Data is summarized in Table 12

Table [ SEQ Table \\* ARABIC ]: Detection Conjugate Titration

Sample Type	25ng/ml		50ng/ml		100ng/ml	
Negative Samples	Mean RLU	CV%	Mean RLU	CV %	Mean RLU	CV%
1	913	20	1828	21	3066	11
2	14572	5	31144	8	52669	10
3	4044	9	8033	7	13339	9
4	13056	6	26847	6	45922	7
5	501	20	1045	35	1184	15
6	561	14	1265	19	2074	16
Mean Negative	5608		11694		19709	
Positive Samples	Mean RLU	CV%	Mean RLU	CV %	Mean RLU	CV%
1	67356	14	135858	18	237651	11
2	268678	8	510962	12	806501	7
3	949499	6	1520675	10	2035020	10
Mean Positive	428511		722498		1026390	
Mean Negative +2 SD	18226		39129		66573	
Modulation (Mean Pos/ Mean Neg+2SD)	24		18		15	

## 2.9 Effect of Sample Dilution[ TC "Effect of Sample dilution" \f C \l "1"

The effect of sample dilution was tested with final sample dilution factors of 1:25, 1:50 and 1:100 PSW into 3% BSA in TBS blocking buffer. Modulation between pooled positive and negative sera was best at 100 fold sample dilution. However, 50 fold sample dilution is also reasonably good. We can observe of a greater reduction in the signal from negative samples compared to the reduction in signal from the positive samples. Results are summarized in Table 13.

**Table [ SEQ Table \\* ARABIC ]: Effect of sample dilution**

Sample Type	25x_PSW		50x_PSW		100x_PSW	
Negative Samples	Mean RLU	CV%	Mean RLU	CV%	Mean RLU	CV%
1	1444	22	660	18	485	12
2	13720	8	6640	24	3475	7
3	3018	14	1924	3	1315	5
4	12214	17	5118	16	3934	8
5	570	23	387	21	328	15
6	537	25	338	24	363	12
Mean Negative	5250		2511		1650	
Positive Samples	Mean RLU	CV%	Mean RLU	CV%	Mean RLU	CV%
1	51798	15	24070	5	12211	11
2	245055	7	132160	8	67462	5
3	889851	13	868038	16	666996	20
Mean Positive	395568		341423		248890	
Mean Negative +2 SD	17377		7940		4927	
Modulation (Mean Pos/ Mean Neg+2SD)	23		43		51	

## 2.10 Effect of changing reagent incubation time [ TC “Effect of changing reagent incubation time” \f C \l "1" ]

The effect of shorter reagent incubation times was tested with sample, detection conjugate and substrate incubation times respectively of 10, 10, 10 and 5, 5, 5 minutes. Eventhough 5,5,5 gave a better modulation than 10, 10, 10 ; it was not chosen because it gave a higher CV%. However, in case of multiplex, 5, 5, 5 can also be an option. There was not much of Assay modulation between the two incubation time and 10, 10, 10 minute incubation protocol was chosen as the final condition (lover CV%).

**Table [ SEQ Table \\* ARABIC ]: Effect of reagent incubation time**

Sample Type	10, 10, 10		5, 5, 5	
Negative Samples	Mean RLU	CV%	Mean RLU	CV%
1	485	12	310	36
2	3475	7	1005	15
3	1315	5	359	14
4	3934	8	1101	4
5	328	15	224	35
6	363	12	238	20
Mean Negative	1650		539	
Positive Samples	Mean RLU	CV%	Mean RLU	CV%
1	12211	11	3462	14
2	67462	5	14914	11
3	666996	20	262970	41
Mean Positive	248890		93782	
Mean Negative +2 SD	4927		1343	
Modulation (Mean Pos/ Mean Neg+2SD)	51		70	

## 2.11 Clinical Sample Correlation and Cut off Determination

Normal donor plasma (N=40) were obtained and tested in the 2 commercial strip kits and in the Theranos System. The Theranos cutoff value was determined by taking the mean RLU of the normal samples plus 5 times the standard deviation of the 40 normal samples (Table 15). The sample RLU divided by the cutoff value yields the Antibody Index. The following criteria was applied to categorize the result as positive (red), negative (green) or borderline (yellow).

Ab Index > 1.1
Ab Index > 0.9, < 1.1
Ab Index < 0.9

**Table [ SEQ Table \\* ARABIC ]: Heterophile antibody assay : Cut off Determination**

Samples	Inter-Cartridge Mean	CV%	Ab Index
1	1309	15	0.14
2	926	10	0.10
3	980	8	0.11
4	458	8	0.05
5	689	9	0.08
6	1274	12	0.14
7	832	16	0.09
8	1150	11	0.13
9	893	22	0.10
10	4029	21	0.44
11	1128	24	0.12
12	1012	22	0.11
13	839	14	0.09
14	584	16	0.06
15	788	17	0.09
16	487	21	0.05
17	3875	15	0.42
18	345	12	0.04
19	1134	14	0.12
20	1048	19	0.11
21	981	19	0.11
22	1813	12	0.20
23	1487	20	0.16
24	436	16	0.05
25	1251	6	0.14
26	934	22	0.10
27	1130	16	0.12
28	1855	16	0.20

[ PAGE \\* MERGEFORMAT ]

29	346	21	0.04
30	760	21	0.08
31	1383	17	0.15
32	926	10	0.10
33	980	8	0.11
34	1347	13	0.15
35	1750	22	0.19
36	2286	17	0.25
37	1529	12	0.17
38	1639	24	0.18
39	8587	11	0.93
40	5487	4	0.60
MEAN	1517		
CUT			
OFF	9186		

Out of the 40 normals tested 39 were negative and 1 was equivocal on the Theranos assay based on the aforementioned cutoff computation. These same samples were all negative except for 1 on one of the 2 Strip kits and the data showed excellent correlation with the Theranos result (Table 16).

For Clinical sample correlation, a total of 58 clinical samples were tested on Theranos assay and on 2 FDA approved commercial kits and the results are summarized in table 16. Excellent correlation was seen for all 58 samples.

**Table [ SEQ Table \\* ARABIC ]: Clinical Samples on Theranos vs. Commercial kits**

<b>Sample ID</b>	<b>Inter-Cartridge Mean</b>	<b>CV%</b>	<b>Theranos Ab Index</b>	<b>Osom Mono Test</b>	<b>Status Mono</b>
1	1309	15	0.14	NEG	NEG
2	926	10	0.10	NEG	NEG
3	980	8	0.11	NEG	NEG
4	458	8	0.05	NEG	NEG
5	689	9	0.08	NEG	NEG
6	1274	12	0.14	NEG	NEG
7	832	16	0.09	NEG	NEG
8	1150	11	0.13	NEG	NEG
9	893	22	0.10	NEG	NEG
10	4029	21	0.44	NEG	NEG
11	1128	24	0.12	NEG	NEG
12	1012	22	0.11	NEG	NEG
13	839	14	0.09	NEG	NEG
14	584	16	0.06	NEG	NEG
15	788	17	0.09	NEG	NEG
16	487	21	0.05	NEG	NEG
17	3875	15	0.42	NEG	NEG
18	345	12	0.04	NEG	NEG
19	1134	14	0.12	NEG	NEG
20	1048	19	0.11	NEG	NEG
21	981	19	0.11	NEG	NEG
22	1813	12	0.20	NEG	NEG
23	1487	20	0.16	NEG	NEG
24	436	16	0.05	NEG	NEG
25	1251	6	0.14	NEG	NEG
26	934	22	0.10	NEG	NEG
27	1130	16	0.12	NEG	NEG
28	1855	16	0.20	NEG	NEG
29	346	21	0.04	NEG	NEG
30	760	21	0.08	NEG	NEG
31	1383	17	0.15	NEG	NEG
32	926	10	0.10	NEG	NEG
33	980	8	0.11	NEG	NEG
34	1347	13	0.15	NEG	NEG

[ PAGE \\* MERGEFORMAT ]

35	1750	22	0.19	NEG	NEG
36	2286	17	0.25	NEG	NEG
37	1529	12	0.17	NEG	NEG
38	1639	24	0.18	NEG	NEG
39	8587	11	0.93	NEG	NEG
40	5487	4	0.60	POS	NEG
41	320948	8	34.94	POS	POS
42	88399	12	9.62	POS	POS
43	143173	11	15.59	POS	POS
44	49020	13	5.34	POS	POS
45	31491	22	3.43	POS	POS
46	14957	8	1.63	POS	POS
47	15771	6	1.72	POS	POS
48	119201	10	12.98	POS	POS
49	156506	10	17.04	POS	POS
50	23409	13	2.55	POS	POS
51	144954	21	15.78	POS	POS
52	121523	11	13.23	POS	POS
53	11028	9	1.20	POS	POS
54	17138	21	1.87	POS	POS
55	32667	4	3.56	POS	POS
56	74602	7	8.12	POS	POS
57	986040	3	107.35	POS	POS
58	502621	9	54.72	POS	POS

## 2.12 Stability Studies

Stability monitoring is ongoing for the the assay reagents stored at 4°C and protected from light for 12 weeks