

Message

From: Ian Gibbons [/O=THERANOS ORGANIZATION/OU=FIRST ADMINISTRATIVE GROUP/CN=RECIPIENTS/CN=IGIBBONS]
Sent: 2/18/2010 7:38:39 PM
To: Sunny Balwani [/O=THERANOS ORGANIZATION/OU=FIRST ADMINISTRATIVE GROUP/CN=RECIPIENTS/CN=Sbalwani]; Elizabeth Holmes [/O=THERANOS ORGANIZATION/OU=FIRST ADMINISTRATIVE GROUP/CN=RECIPIENTS/CN=Eholmes]
CC: Gary Frenzel [/O=THERANOS ORGANIZATION/OU=FIRST ADMINISTRATIVE GROUP/CN=RECIPIENTS/CN=Gfrenzel]
Subject: System 4.0 PPT
Attachments: System 4.0.v2.ppt

As requested ...



System 4.0

System component requirements and selection

02/18/2010

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Overview

System 4.0 will be capable of performing any measurement required *in a distributed test setting*

It is envisaged that *several distinct measurement technologies* will be incorporated

The system will be broadly based on the *existing cartridge and reader concepts*

Open architecture for both reader and disposable

The number of total measurements per sample will be increased by 2 to 3-fold (*target: 15 assays?*)

Assay Menu

- It is crucial to further refine/define which sample types/assays/design requirements are important
- In the field of immunoassay we have good information and experience
- In other fields, we have less secure information
- Specifically:
 - Which nucleic acid analytes?
 - What is the sensitivity requirement?
 - How many and which cell types and surface markers?

Candidate technologies

ELISA using chemiluminescence (current)

ELISA using absorbence

General chemistry using absorbence

PCR and RT-PCR using fluorescence readout

Cell marker assays using laser fluorescence +
movement of detector relative to cells

Electrochemistry for electrolytes and blood gasses

Sample imaging using a camera (see Appendix 3)

General system requirements

- Sample: Blood, plasma and control materials (other types?)
- Sample processing: Plasma from blood, Lysis of cells
- Sample volume: < 20 uL (preferably < 5 uL)
- (For some purposes (e.g. very high multiplex), volumes as high as 200 uL may be permitted.)
- Assay menu: All assays available for system 3.0/3.1 + TBD analytes in other assay classes
- Size, weight: TBD but not larger than System 3.0
- Assay times: TBD but not longer than System 3.1
- Other capabilities: TBD but not less than System 3.0

Proposal for the basis of System 4.0

- Review available technologies: Done
- Define requirements
 - Needs versus wants
- First pass proposal
 - Based on current platform concept (cartridge + dispense)
 - Integrate three optical detection means in one low sample volume device
 - Luminescence
 - Absorbance
 - Fluorescence
 - Add a separate system for cell counting
 - Add red cell removal technology
 - <Add sample integrity evaluation means>
 - Chemistries proposed all exist
- Engineering review

Technology	Analytes	Sensitivity	Dynamic range	Detector Difficulty
Absorbance	1,2,3,4,5,6,7	High	30 fold	Low
Absorbance spectroscopy	1,2,3,4,5,6,7	High	30 fold	Moderate
Turbidimetry	1,3,5	Low	30 fold	Low
Nephelometry	1,3,5	Moderate	100 fold	Low
Fluorescence	1,2,3,4,5,6,7	High	1000 fold	Moderate
Fluorescence spectroscopy	1,2,3,4,5,6,7	High	1000 fold	High
Luminometry	1,2,3,4,5,6,7	V. high	10,000 fold	Low
Cell counting	9	Good	100 fold	Moderate
Cell imaging	9, 10	Moderate	30 fold	High
PCR (and RT-PCR)	8	V. high	10,000 fold	Moderate
Electrochemistry	11, others?	Moderate?	?	Low

Analyte class	Key
Biomarkers	1
Microbial antigens	2
Small molecules (drugs)	3
Small hormones	4
Antibodies	5
Metabolites	6
Enzymes	7
Nucleic acids, viral genomes	8
Cell surface markers	9
Intra cellular markers	10
Electrolytes and blood gases	11

Comparison of Physical Technologies used for Assays

Physical Technology	Application	Mechanical difficulty	Chemistry difficulty	IP/Licensing
Separation	IA	Moderate	Low	
Non-separation	Clinical chemistry	Low	Low	
Non-separation	Nucleic acids	Low	Low	
Non-separation	Immunochemistry	Low	High	Required
Physical multiplex	Any	Moderate	Low	
Chemical multiplex	Immunochemistry	Low	High	
Flow/Movement	Cell counting	Moderate	Low	
Imaging	Cell counting	Low	Low	

Separation of red cells and washing

- There is a trade-off between simplicity of operation of cartridge and instrument versus requirements for (1) low sample volume, (2) using plasma as the sample and (3) washing the capture surface in immunoassays
- We were not able to obtain a sufficient plasma sample from blood for multiplexed immunoassays
- Solutions to this problem are:
 - (1) Use blood as the sample
 - Calibration becomes more complicated
 - (2) Use lysed blood as the sample
 - Calibration should be corrected for HCT
 - (3) Separate plasma *and* use a very low volume measurement technology (e.g. Nanodrop)
 - Will require reverse engineering and/or a license

Special problem for blood samples

Red cell separation means

- Glass fiber wick or pad + plasma extraction
 - Vogel patent has expired
- Magnetic particle agglutination
- ...
- ...
- Pump through a frit??
- ...

Separation versus non-separation technologies

- Theranos has used *separation-based* ELISA as core technology. This requires efficient washing of the assay capture surface to remove unbound label (time consuming).
- There are several immunoassay techniques that have *non-separation* technologies
 - Typically these are:
 - *Proprietary* (license would be needed)
 - Fast
 - Not as sensitive as separation methods
- There are three such technologies that are very sensitive and general
 - EMIT (small molecule drugs) [Syva now Siemens]
 - β -Galactosidase complementation (Biomarkers, receptors etc.) [DiscoverRx]
 - Alpha screen (General immunoassay) [Packard]
- These are reviewed in Appendix 1

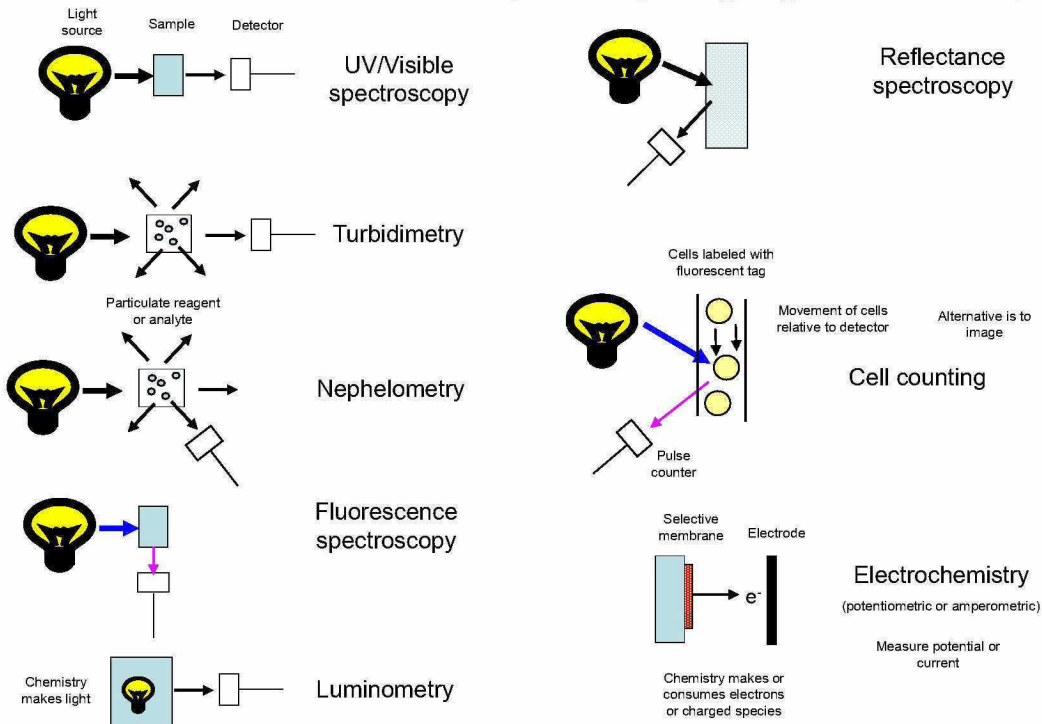
Compatibility and conflict between requirements for the different measurement technologies

- Each distinct detection technology compounds system (instrument and disposable) complexity and cost
 - Is a three-technology system 3x or 9x more difficult to develop?
- Temperature control
 - Thermo-cycler may compromise temperature control for the rest of the instrument
- Light shielding
 - Luminescence detector needs protection from high intensity light sources
- Time factors
 - Current Biomarker assay chemistry takes (say) 30 m
 - RT-PCR may take > 2 hours

Trade-off of sensitivity and time

- Luminescence ELISA can generally be complete in 15-30 m
- High sensitivity absorbance ELISA may need hours for equivalent sensitivity
- PCR sensitivity increases with # of cycles
 - 10 cycles: 10^3 -fold
 - 20 cycles: 10^6 -fold
 - 30 cycles: 10^9 fold

Detection configurations compared (imaging not shown)



Requirements, capabilities and limitations of candidate detection technologies and chemistries

ELISA with Chemiluminescence readout

- Very sensitive with very wide dynamic range
- Capable of giving results rapidly
- Suitable for *immunoassay* of small and large analytes
- Weakness is that reagent stability (E-Ab) and substrate contamination are crucial

- **Essential technology**

Absorbance

- **Many applications**
 - General chemistry (e.g. cholesterol)
 - Hemoglobin (and calculated HCT)
 - Albumin and total protein
 - TDM by licensed assays (EMIT/DiscoverX etc.)
 - Enzyme assays (e.g. ALT)
 - ELISA with absorbance readout
 - Enables adaptation of existing assays?
 - Electrolytes (e.g. Ca²⁺)
 - Sensitive with moderate dynamic range
 - Take longer than those with chemiluminescent readout
- **For full value wavelength range of 340 – 650 needed**
- **Essential technology**

Fluorescence

- **Essential** for nucleic acid assays
- Not needed for clinical chemistry or immunochemistry
- Good sensitivity and dynamic range
- Capable of multiplexing

- Requires a high quality light source (laser[s])
- Compensation for light source instability is needed

PCR and Reverse Transcriptase-PCR

- **Essential** for all nucleic acid assay targets
- Requires both elevated temperatures (90C) and temperature cycling
- Likely to require development of new disposable elements/surface chemistry
- Needs fluorescent readout

Cell counting

- Questions:
- How many markers?
 - >> How many lasers and complexity of the optics
 - One laser gives three colors
- Is rare cell detection needed?
 - Problem is sample volume and the need for cell concentration prior to detection

Rare cell analysis

- Circulating cancer cells
- Fetal cells in maternal blood

- **Problem:**
 - Cell numbers may be as low as (say) 10/mL
 - Blood drop = 20 uL
 - Cells/drop = 0.2: Impossible to detect reliably
- **Solution is to pre-concentrate target cells from (say) 1 mL of blood then to detect**
 - Not compatible with POC context
 - Technically demanding



Technologies deemed unnecessary

Reflectance

Imaging

Electrochemistry



theranos
redefining healthcare

Attractive potential add-on technologies (See appendix 1)

- Non-separation, receptor-based assays with absorbance or luminescence readout
- Pulse oximetry

Review of issues for technologies not yet familiar at Theranos

Nucleic acid assays

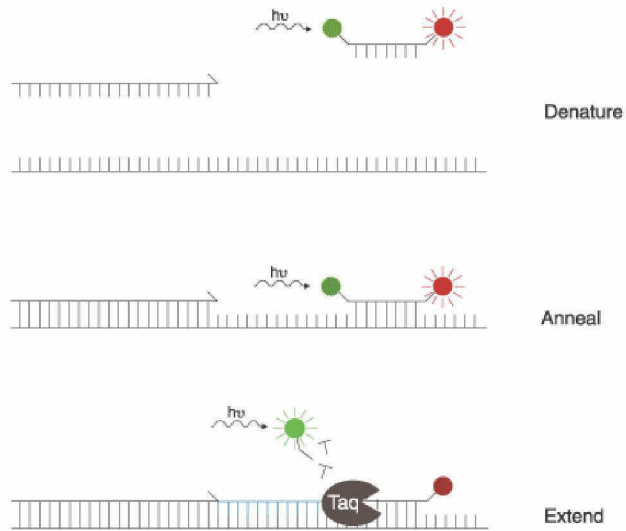
- **Polymerase Chain reaction (PCR)**
 - Amplify DNA: 30 cycles = 10^9 -fold amplification
 - Needed for sensitivity
- **Reverse transcriptase (RT-PCR)**
 - Measure RNA (transcribe to DNA then apply PCR)
 - Needed for RNA virus detection
- **Real time PCR**
 - Semi quantitative
- All need temperature cycling and elevated temperatures
- Read-out by fluorescence

PCR and RT-PCR:

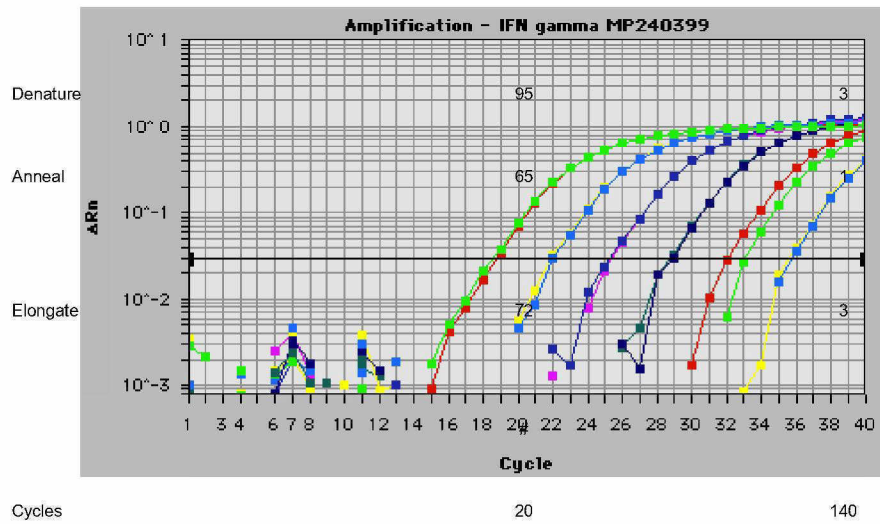
Time and temperature requirements, typical

Step	Temperature, C	Time, min
Denature	95	3
Anneal	65	1
Elongate	72	3
Cycle	#	
	20 (10 ⁶ fold amplification)	140

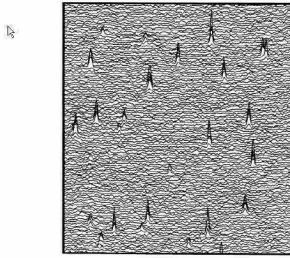
Real-time PCR (TaqMan) One favored readout means (needs a license)



Real-time PCR read-out



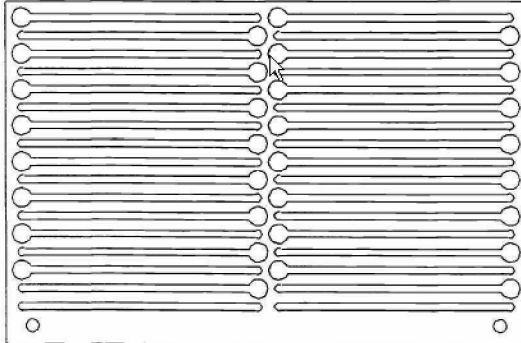
Cell counting: option 1



Microvolume Laser Scanning Cytometry Platform for Biological Marker Discovery

Ian D. Walton, Louis J. Dietz, Gary Frenzel, Jerry Chen, Jim Winkler, Scott M. Norton,
Aaron B. Kantor

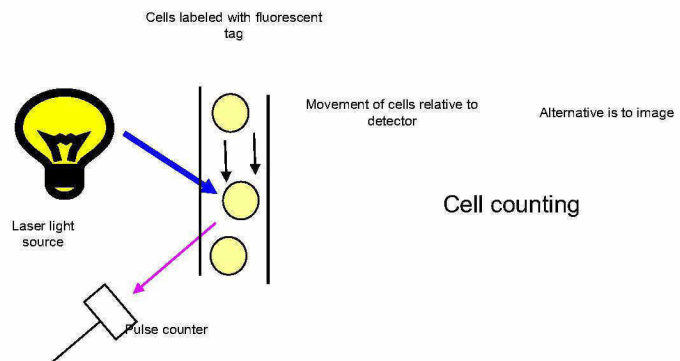
SurroMed Inc., 1060 East Meadow Circle, Palo Alto, CA 94303.



- Spatial array of stained cells
- Raster optics (move laser)
- Or, image
- Simple capillary cartridge (shown for analysis of 32 samples)

Cell counting: Option 2

- Needs controlled flow
- Measure pulses over *time*



Questions/Issues

Complexity versus cost and practicality

Nanodrop does not measure absorbance and fluorescence in the same unit

Laser: size/cost versus # of cell surface markers

Costs

Do we need electrochemistry?

- Many analytes measured by electrochemical methods (O_2 , CO_2 , HCO_3^- , Ca^{2+} , Mg^{2+}) are only needed in emergency situations
- Many analytes such as K^+ , Ca^{2+} , Mg^{2+} , Phosphorus (PO_4^{2-}) and pH can be measured by absorbance
 - Protocols are simple
 - Two or three liquid reagents
 - Mix with sample and incubate



Small volume assay reader concept

02/18/2010

Development Team

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theranos
redefining healthcare

Strategic Issues For POC Assay Systems

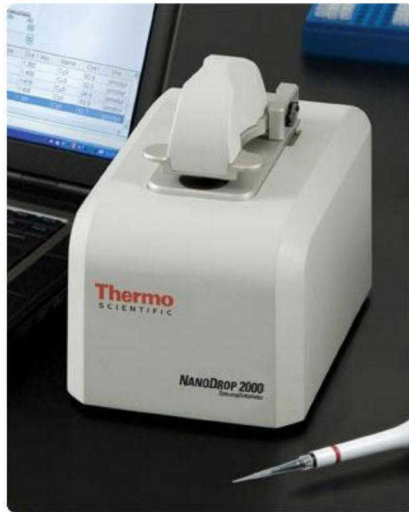
- Performance equivalent to laboratory methods
 - Multiplexed assay capability
 - *Multiple assay type capability*
 - *Immunoassays*
 - *Direct assays (cholesterol etc.)*
 - *Enzyme assays*
 - *Nucleic acid assays*
 - *Electrolytes (and blood gasses??)*
 - **Simplicity and reliability of use**
 - Non-technical users
 - No false results
 - *Small sample volume*
 - *< (say) 5 uL blood*
 - *No hematocrit effect*
 - **Speed**
 - *< (say) 15 m*
 - *Ease of assay development*
 - *Speed of assay development*
 - **Low cost**
 - *Instrument*
 - *Disposable*
- Italics emphasize possible improvements/extensions on/of current system



Small volume assay concept

- Blood sample: say 10 uL (= about 5 uL plasma)
- Sample capillary sucks plasma (say 3 uL; note filter does not need to be very efficient) from a (say) glass fiber filter.
 - Could also use a frit in a "tip" (?) + aspiration
- Plasma fills capillary and is then displaced into a dilution well. Mixing by repeated aspiration and re-expression >> (say) 30 uL diluted plasma
- Diluted sample (3 uL) is aspirated into standard Theranos tips (up to 8 assays = 24 uL diluted sample)
- Process assays with color forming chemistry (ELISA) generating (say) 3 uL colored product
 - Tip precludes evaporation
- Read in Nanodrop-style photometer
- Advantages
 - HRP chemistry enabled
 - Small sample volume
 - Red cell removal would be possible
 - Higher multiplex might be possible
 - Can also be used for clinical chemistry assays
 - Glucose, Cholesterol
 - Enzymes (ALT)
- Disadvantages
 - Light source required (can be very simple and inexpensive as can the detector, however)
 - Need to wash sample chamber (Nanodrop cleans up fine by just wiping +/- one wash)

“Nanodrop” spectrometer



- Designed to measure protein and nucleic acid concentrations in very small volumes
- 200 – 800 nm
- 0.5 – 5 μ L
- Measurement complete in a few seconds

Use of Nanodrop



Pipette (say) 2 μ L
onto pedestal



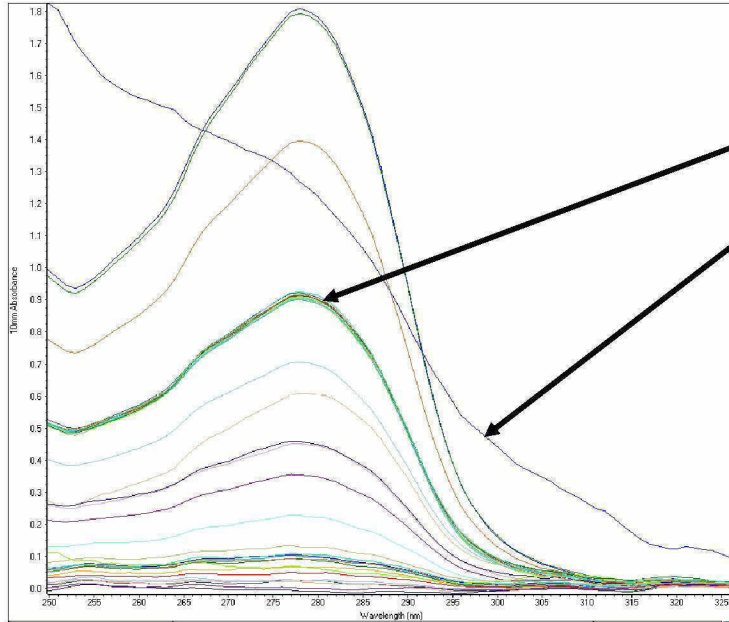
Instrument automatically
defines a precise optical
pathlength (1 mm)

Instrument Specifications

NanoDrop 2000/2000c – pedestal mode

Instrument Type:	Spectrophotometer
Minimum Sample Size:	0.5 μ L
Pathlength:	1 mm (auto-ranging to 0.05 mm)
Light Source:	Xenon flash lamp
Detector Type:	2048-element linear silicon CCD array
Wavelength Range:	190-840 nm
Wavelength Accuracy:	\pm 1 nm
Spectral Resolution:	\leq 1.8 nm (FWHM @Hg 253.7 nm)
Absorbance Precision:	0.002 absorbance (1 mm path)
Absorbance Accuracy:	\pm 2% (at 0.76 absorbance at 257 nm)
Absorbance Range:	0.02 -300 (10 mm equivalent)
Detection limit:	2 ng/ μ L dsDNA
Maximum Concentration:	15,000 ng/ μ L (dsDNA)
Measurement Time:	< 5 seconds
Footprint:	14 cm x 20 cm
Weight:	2.0 kg
Sample pedestal Material of Construction:	303 stainless steel and quartz fiber
Operating Voltage:	12 VDC
Operating Power Consumption:	12-18 W, (max 30 W)
Software Compatibility:	Windows® XP and Vista (32 bit)

Protein spectra



- Serial dilutions of a protein solution are shown
- Measurements are reproducible
- Spectral ratios make it easy to detect malfunctions (e.g. bubbles in the light-path)

Nanodrop applications illustrated

Performance measures

Sample volume independence

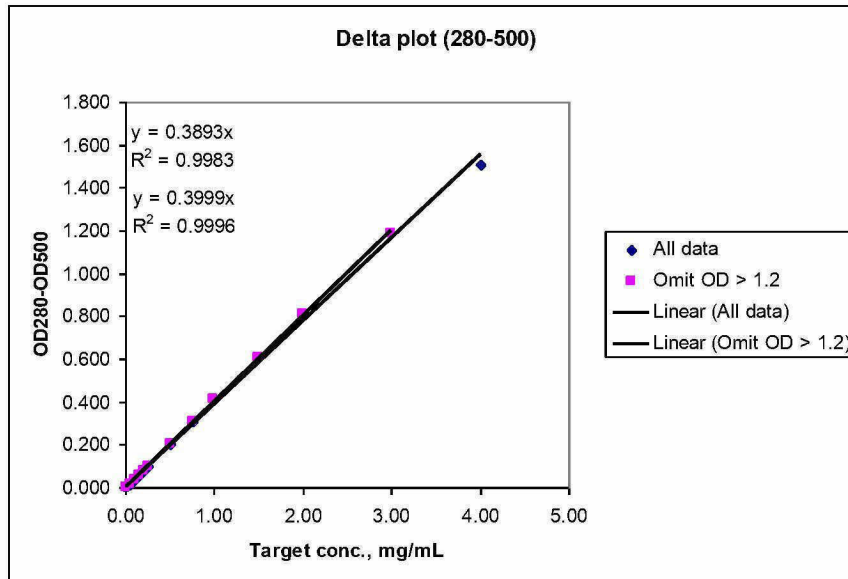
Protein assay (UV measurements)

Measurements in the visible

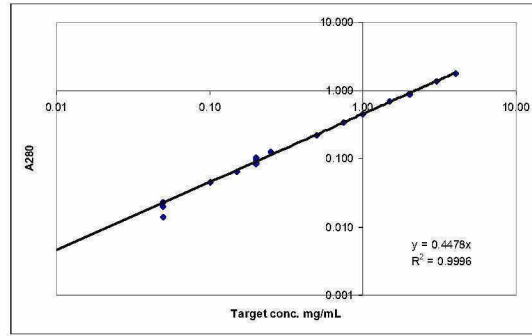
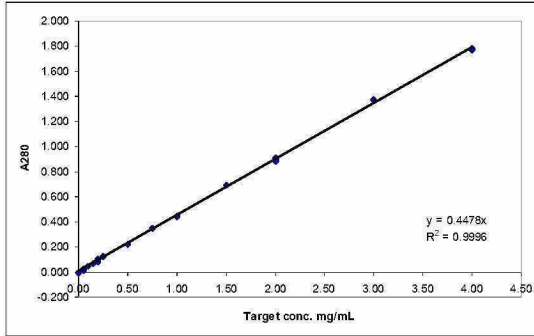
ELISA readout

Hemoglobin/HCT measurement

Dose-response is linear up to about 1.5 OD



Good linearity from low to high absorbance

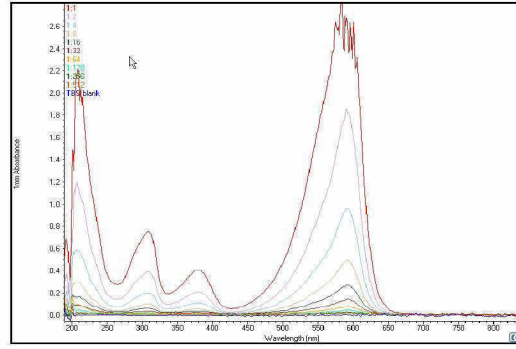
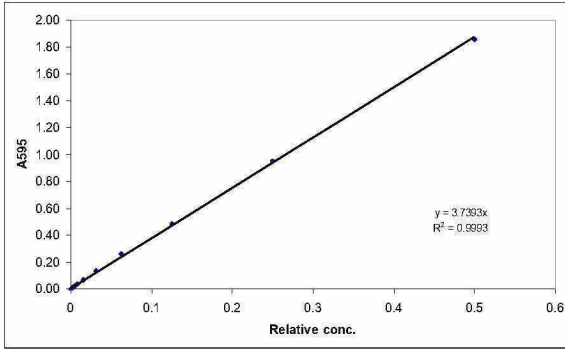


Precision is good. Even at low A values

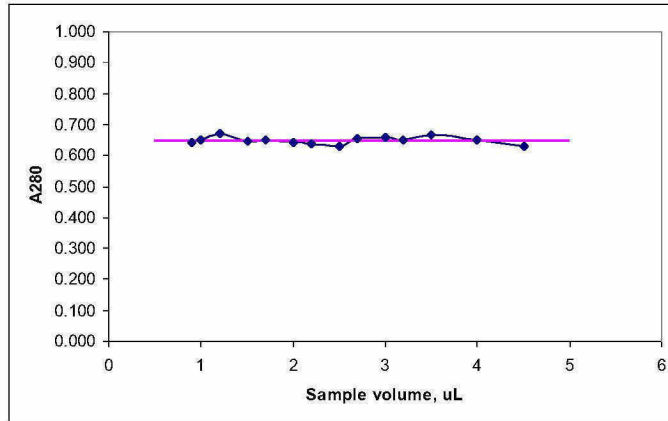
BSA, mg/mL nominal	A, 1cm,280, avg	N	CV, %
2.0	0.923	11	0.9
0.2	0.093	5	8.2
0.05	0.019	4	18.6

LOD (95% confidence) = 0.01 (Absorbance); 0.02 mg/mL (BSA conc.)

System also works in the visible range

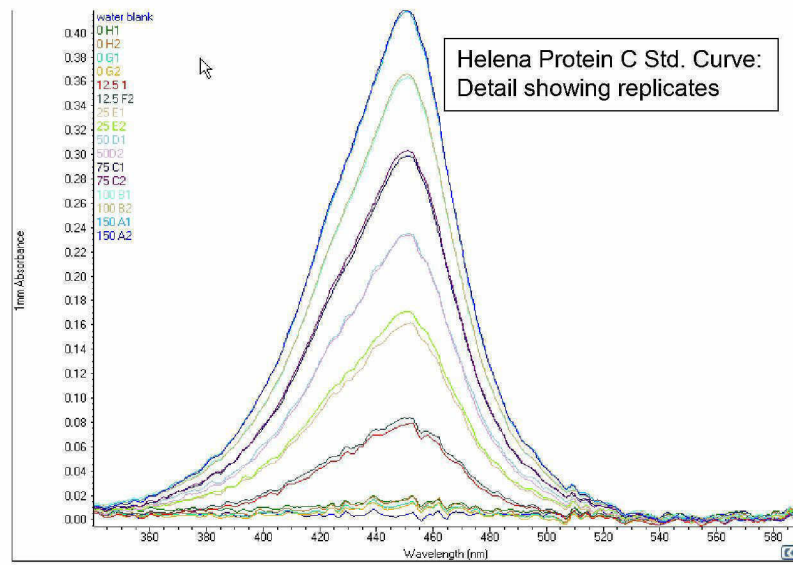


System response is independent of sample volume

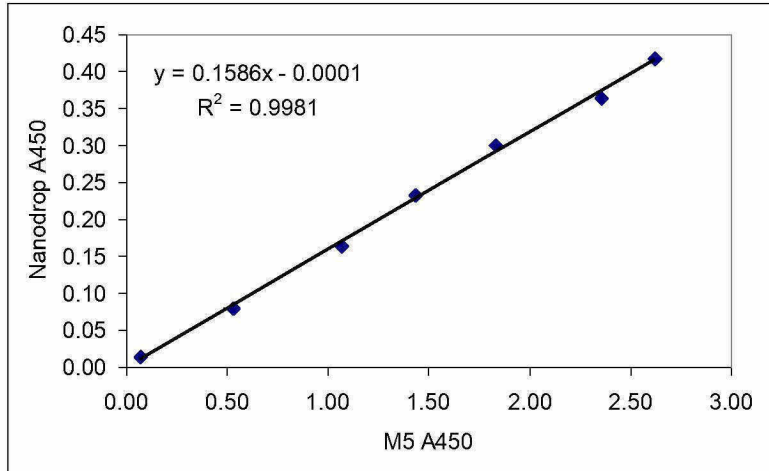


average	0.648
sd	0.011
cv, %	1.744

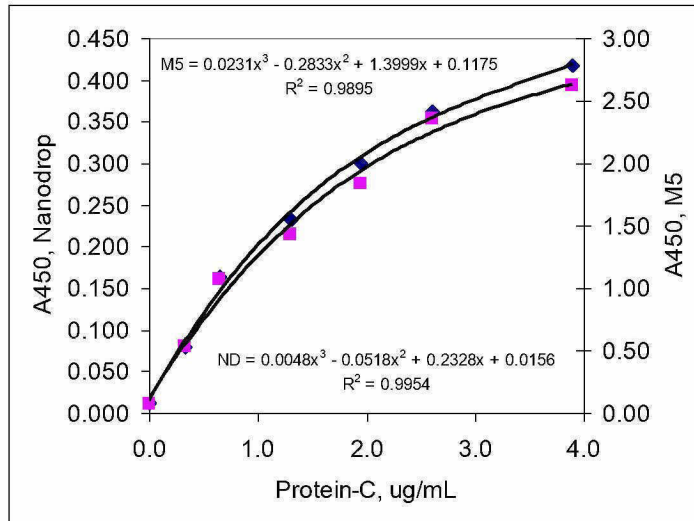
ELISA: Visible spectra



ELISA assay comparison (signal) Helena Protein-C assay

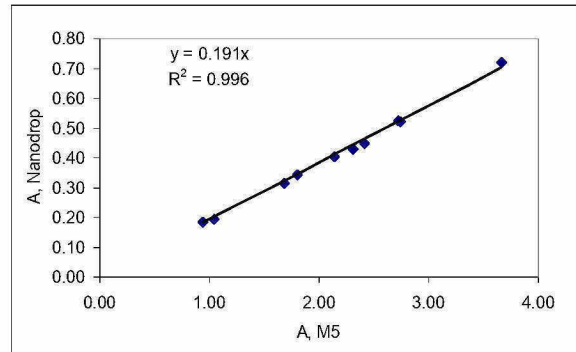
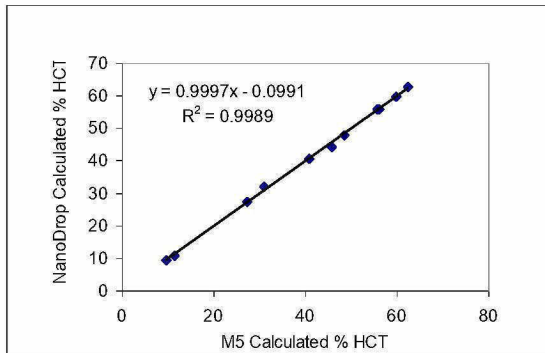


Protein-C ELISA response comparison



Hematocrit/Hemoglobin assay

Standard clinical assay for Hb (Drabkin's method)
(RBCs are lysed; Hb is converted to Cyan:Met-HB; read A540 nm)



Possible applications in a next generation Theranos Instrument

- Colorimetric ELISAs
 - HRP or APase labels
- Clinical chemistry analytes can be measured colorimetrically
 - Glucose
 - Cholesterol
 - HDL-cholesterol
 - Electrolytes
- Enzyme and other types of assays with NADH or NADPH readout
 - EMIT! (TDM Assays!)

Signal is lower than for a standard spectrometer

- Path length is 1 mm
- A is about 10 – 20 % that of a MTP reader (path length 0.5 – 1.0 cm)
- ΔA (SD) 0.002 *i.e.* CV = 10% at A = 0.02 (OD 0.2)
- This can/may be compensated for by letting signal (A) values rise into the range (say) 0.2 – 5.0
 - Elisa assays: let enzyme work longer and use elevated temperature
 - General Chemistry: dilute less

Nanodrop 3300: Spectrofluorometer



Q: Is the Thermo Scientific NanoDrop 3300 an upgrade to the NanoDrop™ 2000?

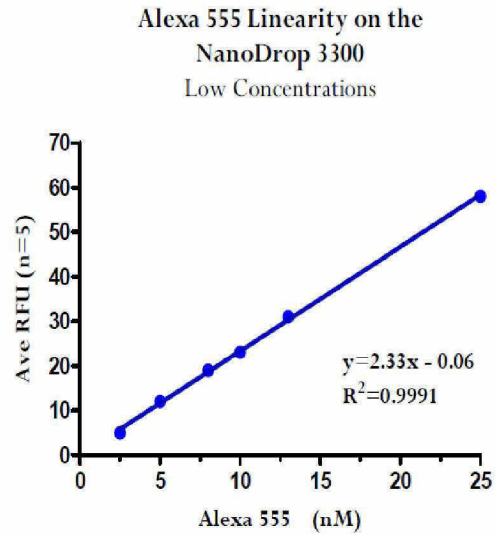
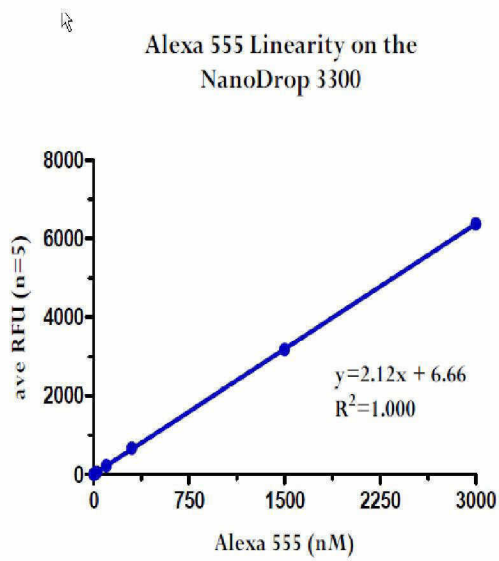
A: No, the NanoDrop 3300 is used to measure fluorescence whereas the NanoDrop 2000 is used to measure absorbance.

Nanodrop Fluorimeter Specifications

Specifications

Instrument Type	Fluorospectrometer
Minimum Sample Size	1 µl
Light Sources	3 light emitting diodes (LEDs)
Excitation Maxima	UV: 365 nm Blue: 470 nm White 460-650 nm
Detector Type	2048 - element linear silicon CCD array
Wavelength Range	400-750 nm
Wavelength Accuracy	1 nm
Spectral resolution	8 nm (FWHM at Hg 546 nm)
Fluorescence Precision	< 5% CV (10 nM fluorescein)
Detection Limit	< 1 fmol fluorescein
Measurement Cycle Time	2 - 10 seconds
Dimensions (footprint)	14 cm X 20 cm
Weight	1.5 kg
Sample Pedestal Material of Construction	303 stainless steel and quartz fiber
Operating Voltage	5 vdc (supplied by USB port, no external power supply)
Operating Power Consumption	2 W
Standby Power Consumption	1 W
Software Compatibility	Microsoft Windows® 2000, XP, and Vista (32 bit)
UL/CSA and CE	All units are approved to these standards

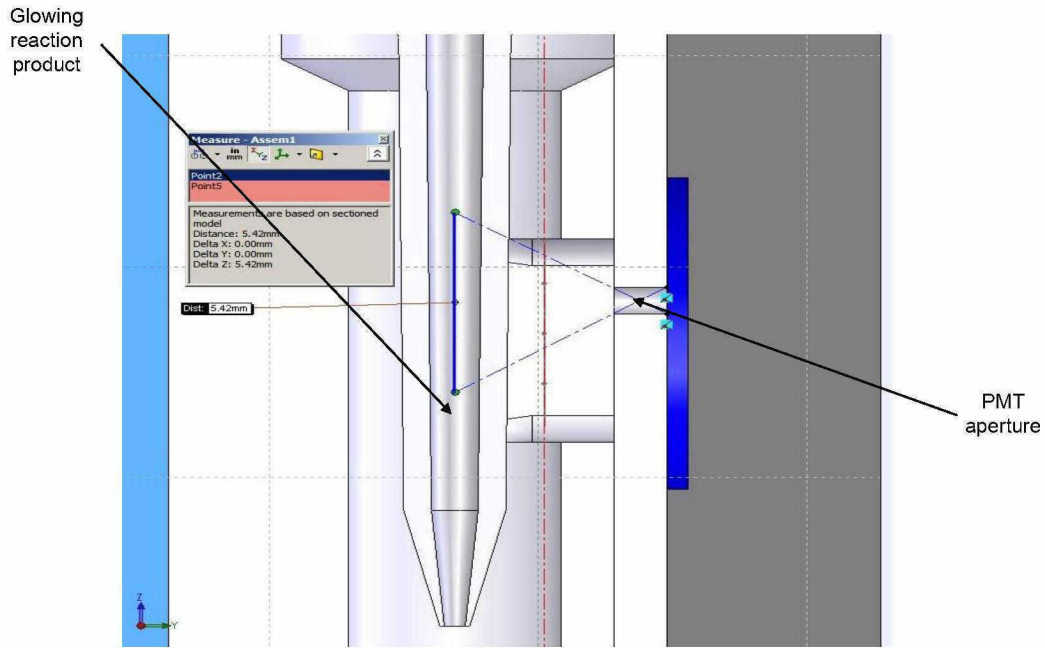
Nanodrop Fluorimeter: Sensitivity and dynamic range



Question and Idea

- Why not adapt the Nanodrop spectrometer to chemiluminescence?
- Develop a multi-detection mode platform
 - Absorbance
 - Fluorescence
 - Luminescence
- Problem might be smaller volume >> less signal
- The Nanodrop optic, however, appears to use a larger fraction of the sample volume than conventional spectrometers (and the Edison?)
 - Edison light collection volume has been estimated as about 5 μ L but only a fraction of the light from that volume gets to the PMT.

Current Instrument Light Collection



US 6,628,382 Sept. 30 2003 CW Robertson

Priority date: 08/1999

1. A photometric or spectrophotometric apparatus wherein a sample in the form of a liquid drop is contained by surface tension forces between two planar surfaces, one containing a photometric or spectrophotometric source and the other a photometric or spectrophotometric detector and an optical path is established through the sample between the two surfaces said apparatus comprising:

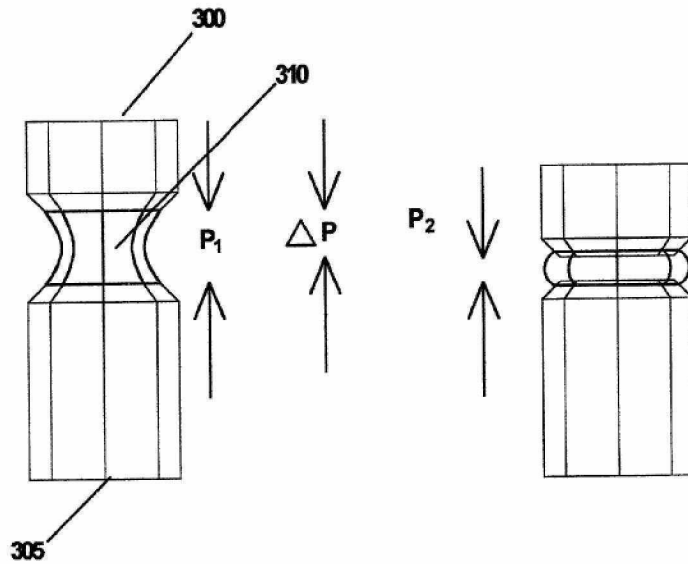
first and second anvil surfaces at least one being moveable relative the other to any one of three positions:

an adjustable sample loading position so selected that the at least one moveable surface and the other surface are so remotely spaced that a liquid drop can be placed on the first surface;

an adjustable sample compression position so selected that the surfaces are opposed and substantially parallel and proximally spaced so that the liquid wets and spreads upon both surfaces;

an adjustable sample measuring position so selected that the opposed substantially parallel surfaces are spaced apart to pull the sample into a column wherein it is contained by surface tension thereby providing an optical path for a photometric or spectrophotometric measurement.

NanoDrop: Definition of light path

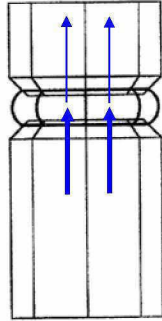


Sample pathlength and illuminated volume are defined by the geometry of the pedestal

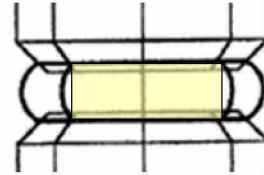
Volume in excess over 1 uL is expressed out of the light beam and contained by surface tension

Optics For Three Detection Technologies (1) Absorption

Detector

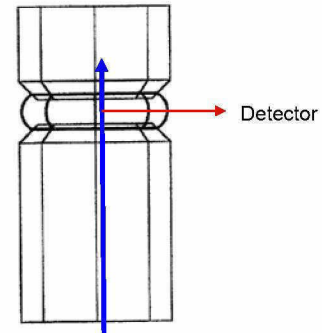


Light source producing
a wide highly-collimated
beam

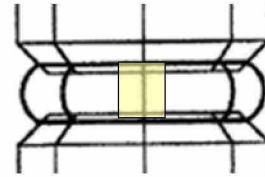


Defined illuminated
volume

Optics For Three Detection Technologies (2) Fluorescence



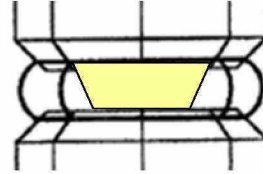
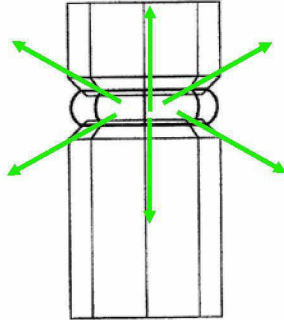
Light source producing
a *narrow* highly-
collimated beam



Defined illuminated
volume

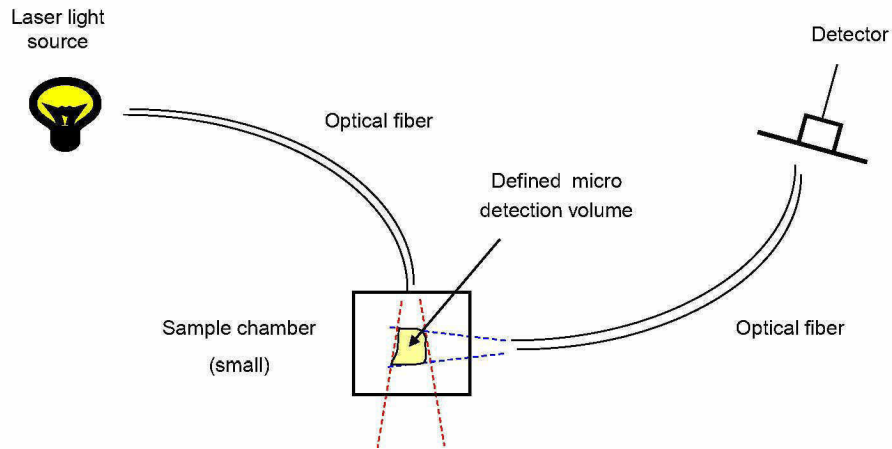
Optics For Three Detection Technologies (3) Luminescence

Detector



Defined volume for
light collection

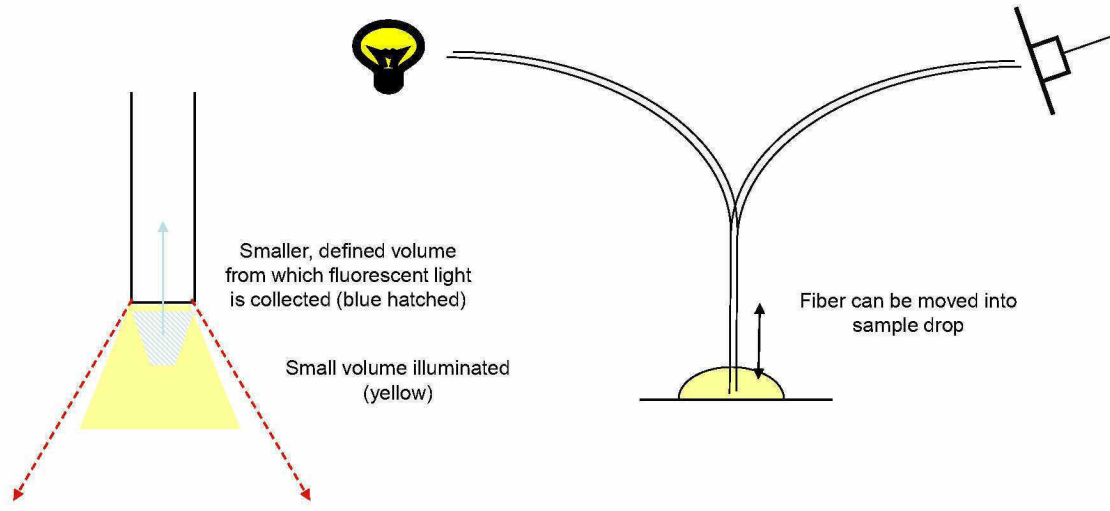
Alternative low volume system for optical detection of fluorescence using optical fibers



See also: US 4,676,640 (expired)

[US4676640.pdf](#)

Another version of the fiber approach for low volume fluorescence detection and cell counting



See: US 4,676,640 (expired)

[US4676640.pdf](#)

What would be needed to incorporate this technology in our system?

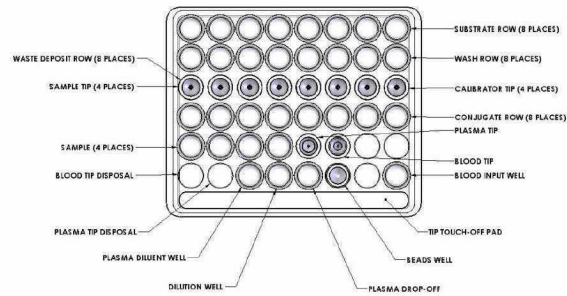
- Interface with liquid dispensing
 - Demo in progress
- Interface with RBC filtration
- Add a simple light source to the instrument
- Detector
 - PMT? or simple CCD (spectroscopy [many wavelengths]) or Photodiode (one color)
- IP: license or (better) reverse engineer

Recommendations

- Selected candidate technologies
 - Luminescence
 - Absorbance
 - Spectroscopy needed (multi-wavelength)
 - Wavelength range 340-650 nm
 - Fluorescence
 - RT-PCR
 - Cell counting
- Need to *consolidate* optics around a low volume detection means (*invention needed*)
 - Nanodrop-like?
- Need to *invent* and develop red cell separation technology

Pipettor head multiplicity: Issues

- **Holding liquid during incubations**
 - “Tip” versus “well”
 - Gravity
 - Tip orientation
 - Small volume would likely remain fixed (even if tip is vertical)
 - Evaporation (very important if volume < 5 uL)
- **Pressure transmission when picking up tip**
 - Premature ejaculation
- **Precise positioning**
 - Nanodrop approach would require good precision in x, y and z
 - < 0.1 mm is OK and easy to implement
 - But multiple head compounds the problem
- **Temperature control**
 - Absolute requirement for immunoassays and PCR



- Current design completely fills the footprint (close packing of assay elements)
- Footprint is dominated by *diameter* of reagent tubes and the tip boss
- Can these be smaller?
- Can some reagents be shared?
 - One head concept makes this easier
 - But, there is a time penalty

Engineering evaluation

- **Feasibility review**
 - Complete analyte list
 - Complete chemistry review against all analytes
 - Are integrated optics feasible?
 - How to implement more assays on same footprint
 - Can footprint be bigger?
 - Cartridge height could be less for smaller volume assays
- **Development plan**
- **Costs and Development Time**

Appendix 1

Some alternative technologies including non-separation methodologies

Review of Theranos requirements against possible licensable technologies



Technology Review

April 2009

02/18/2010

This presentation and its contents are Theranos proprietary and confidential



Theranos mission Instrumented system

To enable rapid, accurate, multiplexed POC
measurements of key biomarkers and therapeutic
agents

To replace laboratory methods in monitoring disease
and therapy

Capabilities (ideal)

Sample types

- Blood (F/S, venous), serum, plasma, urine (?)

Marker types

- Proteins
- Small molecules
- Metabolites
- Nucleic acids
- Cell markers (Surface + Internal)
- Electrolytes

Number of multiplexed markers

- Up to 20

Number of assays available

- >100

Specificity, accuracy, precision and sensitivity

- Match current state of the art

Size and speed

- Match current POC state-of-art
 - Time < 15 min

Stability

- > One year at RT

Theranos cost per result

- < \$3 equivalent to \$18/six plex; Price = > \$100 at > 80% gross margin

Disposables/year

- > 1,000,000

FDA, CLIA cleared, ISO-9xxx

Patent protected

Sample integrity and volume measurement

Sample pre-treatment (including removal of red cells)

Sample types	Match
<ul style="list-style-type: none"> • Blood (F/S, venous), serum, plasma, urine 	
Marker types	Match
<ul style="list-style-type: none"> • Proteins • Small molecules • Metabolites • NOT Nucleic acids • Cell markers (Surface + Internal) 	Better Match Worse N/A Worse
Number of multiplexed markers	Worse (30-plex available)
<ul style="list-style-type: none"> • Up to six 	
Number of assays available	Worse
<ul style="list-style-type: none"> • 100 	
Specificity, accuracy, precision and sensitivity	Match
<ul style="list-style-type: none"> • Match current state of the art 	
Size and speed	Worse
<ul style="list-style-type: none"> • Match current POC state-of-art <ul style="list-style-type: none"> • Time < 15 min 	
Stability	Worse
<ul style="list-style-type: none"> • > One year at RT 	
Theranos cost per result	?
<ul style="list-style-type: none"> • < \$3 - equivalent to \$18/six plex; • Price => \$100 at > 80% gross margin 	
Disposables/year	?
<ul style="list-style-type: none"> • > 1,000,000 	
FDA cleared	Worse
Patent protected	Worse (Situation improving this year)



Current capabilities

<u>Attribute</u>	<u>Current</u>	<u>Possible</u>
Sample types	YES	
• Blood (F/S, venous), serum, plasma, urine		
Marker types		
• Proteins	YES	
• Small molecules	YES	
• Metabolites	NO	YES
• Ions (Ca ²⁺ etc)	NO	?
• NOT Nucleic acids	N/A	?
• Cell markers	NO	? (NEW SYSTEM)
Number of multiplexed markers	NO	YES
• Up to six		
Number of assays available	NO	YES
• 100		
Specificity, precision and sensitivity	YES	
• Match current state of the art		
Size and speed	NO	MAYBE
• Match current POC state-of-art		
• Time < 15 min		
Stability	NO	YES
• > One year		
Theranos cost per result	NO	MAYBE
• < \$3 equivalent to \$18/six plex;		
• Price = > \$100 at > 80% gross margin		
Disposables/year	NO	YES
• > 1,000,000		
Instruments/year	YES	
• 1,000		
FDA, CLIA etc. cleared	NO	YES
Patent protected	NO	YES

Existing technology capabilities

Sensitivity limit (Can we do better for biomarkers? NO)

Calculations	Chemiluminescence EIA	
Specific signal		
Antigen conc.	5.00E-01 pg/mL	Current best
Dilution	3.00E+00 fold	
Diluted antigen conc.	1.67E-01 pg/mL	
Efficiency of binding	1.00E+01 %	
Enzyme molecules/antigen	2.00E+00 mole/mole	Guess
Enzyme turnover	5.00E+04 sec ⁻¹	
Enzyme conc.	3.33E-02 pg/mL	Apase
Interogated volume	1.00E+00 uL	
Enzyme quantity	3.33E-05 pg	Only see part of liquid column
Enzyme MW	1.20E+05	
Avogadro's #	6.04E+23	
Vmax	8.39E+06 molecules of product/sec	
S/K	5.00E-02	
v	4.19E+05 molecules of product/sec	Substrate is not saturating
Quantum yield	5.00E-01 %	
Photon production	2.10E+03 counts/sec	Guess
Detection efficiency	5.00E+01 %	
Signal	1.05E+03 counts/sec	
Enzyme molecules	1.68E+02	
Background signal		
Signal	100 counts/sec	
Signal	300 counts/sec	Substrate blank
S/B	5.59E-01	NSB (best case)

The clinical laboratory ***

Service providers ***

Platforms for customer assay development

ELISA

- Multiplexed Imaging (Searchlight)

Lateral flow (Biosite)

Electrochemiluminescence (MSD)

Time resolved fluorescence (TRF)

Non-separation methods

- Alpha screen (Packard)
- Enzyme complementation (DiscoverRx)
- Homogeneous time-resolved fluorescence (HTRF)
- EMIT (small molecules)
- FP (small molecules)
- Surface plasmon resonance ("Reagent-free", Biacor, Forte Bio)
- Evanescent wave

FACS screen

- Guava

Bead-based multiplexes (Luminex, BD etc.)

POC niche

- Cholestech (Lipids)
- Abaxis (General chemistry + low sensitivity immunoassay)
- Coagucheck etc. (PT)
- Hemocue (Hemoglobin)
- Many (blood glucose)
- Metrika (HbA1c)
- Ions (ISE) + IA (I-Stat)

Factory calibration

Validation problems (multiplexes)

Sample handling (blood)

- Stability

Sample volume (blood)

- Too large



Potential improvements to our existing system

Up to 12 assays

Smaller sample 10 uL at 6-plex?

Faster (assays complete in < 15min?)

Smaller (1/4 th current size?)

More precise (< 5 %CV?)

More reliable (< 1/1000 failures?)

Add selected general chemistries

Add other types of marker

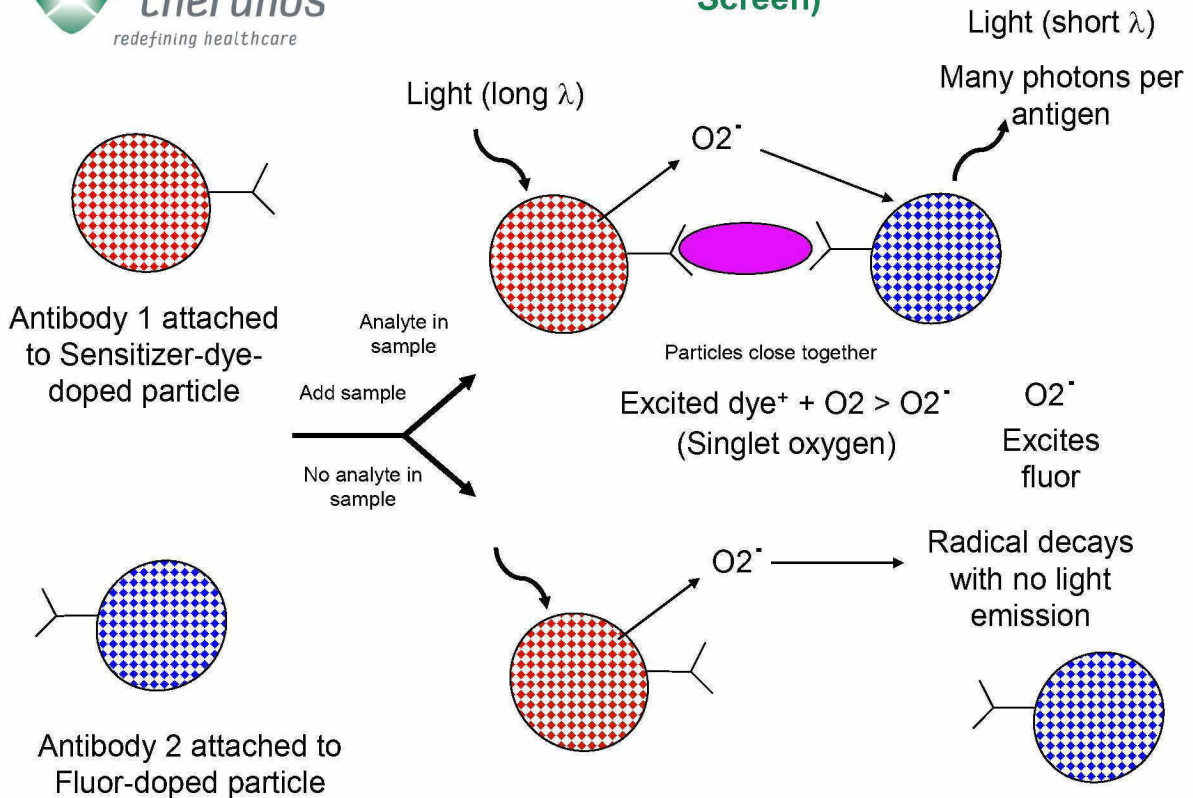


Candidate supplemental/alternate technologies

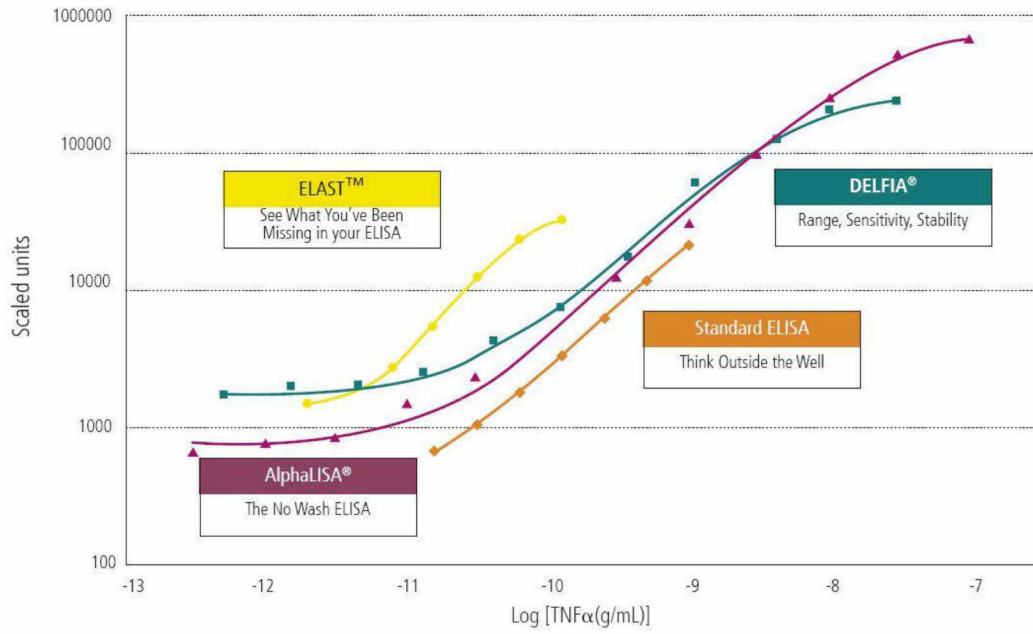
Examples

- DiscoverX
- AlphaLISA
- EMIT
- HTRF
- ELAST
- ...

Luminescence Oxygen Channeling IA (LOCI, Alpha Screen)



Technology comparison



Enzyme complementation (DiscoverRx) β -Galactosidase

Native β -Galactosidase polypeptide



Mutant, inactive β -Galactosidase polypeptide (EA)



Small N-terminal polypeptide (ED)



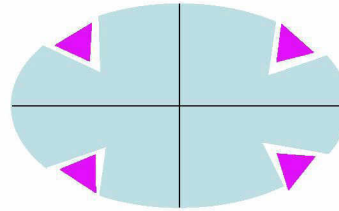
Enzyme Donor "ED" (Inactive)



Mix



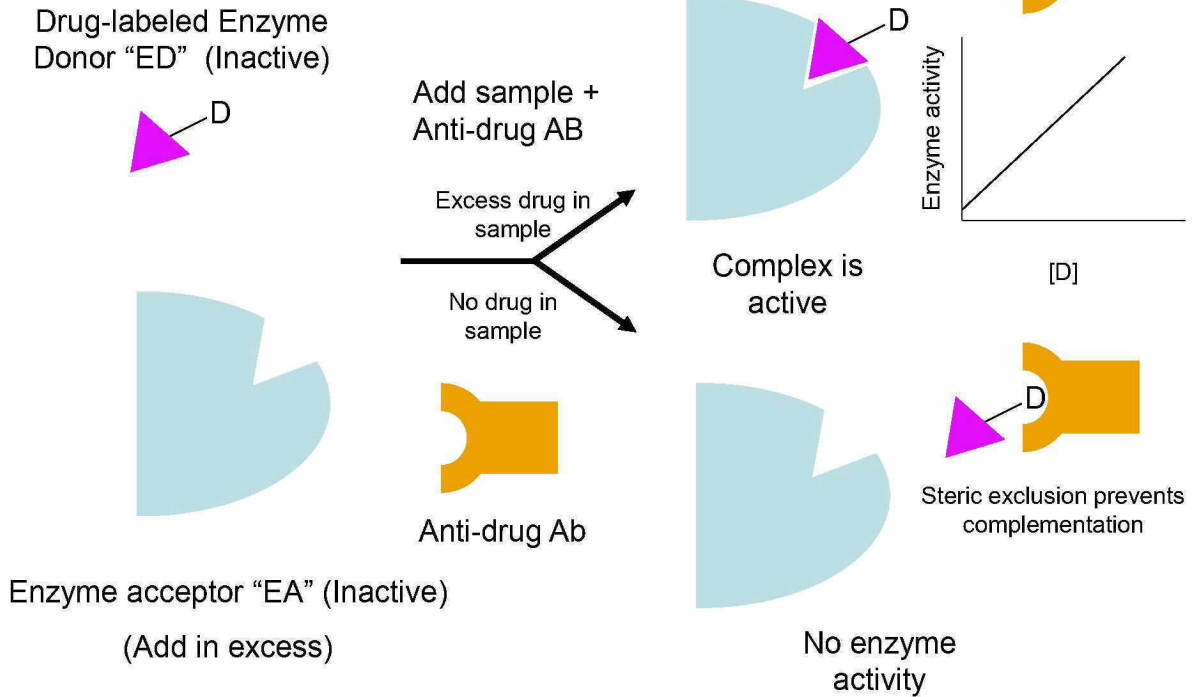
Native enzyme is a tetramer



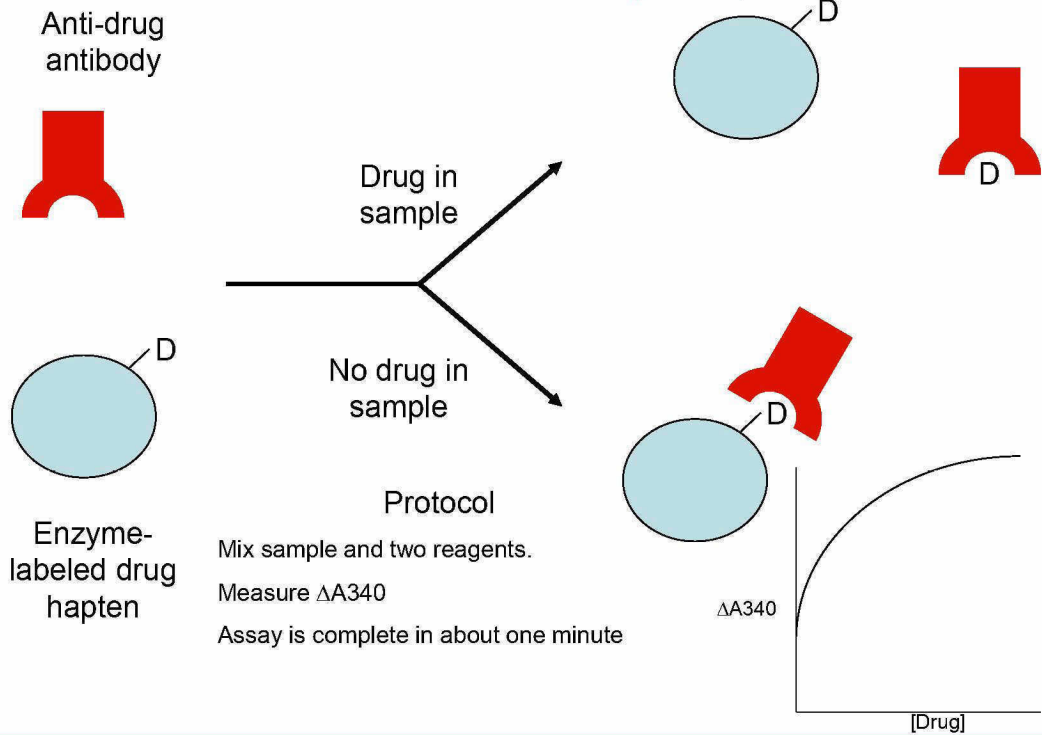
Complex (Active) forms quickly

Enzyme acceptor "EA" (Inactive)

Enzyme complementation immunoassay (Non-separation, rapid)



EMIT (Enzyme multiplied immunoassay technique)
Rapid, non-separation measurement of small molecule drugs
(Siemens)



- “Homogeneous time-resolved fluorometry”
- No separation required

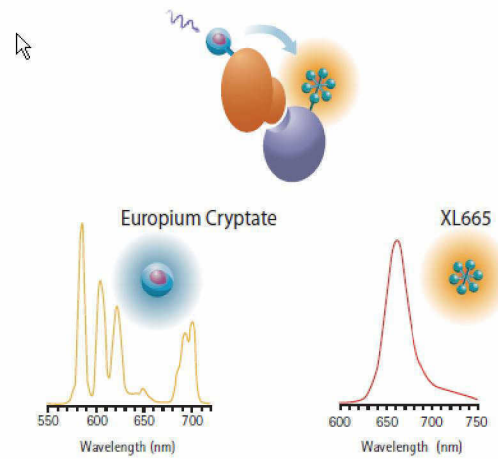


Fig.1 : HTRF principle with Eu³⁺ cryptate and XL665 as respectively donor and acceptor. When the two entities come into close proximity and upon excitation, FRET occurs and XL665 re-emits a specific long-lived fluorescence at 665 nm.

- Dissociation-enhanced lanthanide fluorescence immunoassay
- Europium chelate labeling
- Fluorescence enhanced by release of Eu and shielding on a micelle

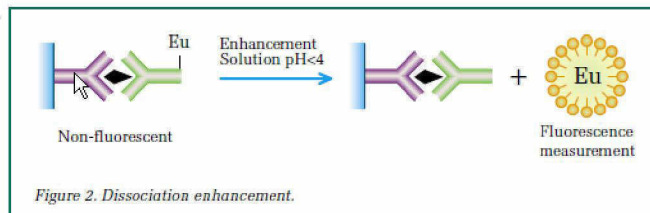
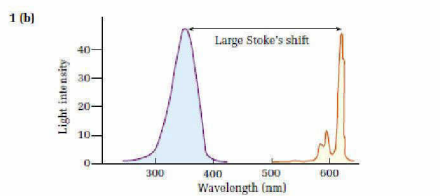
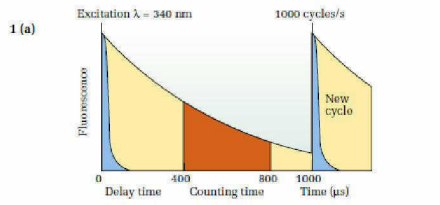
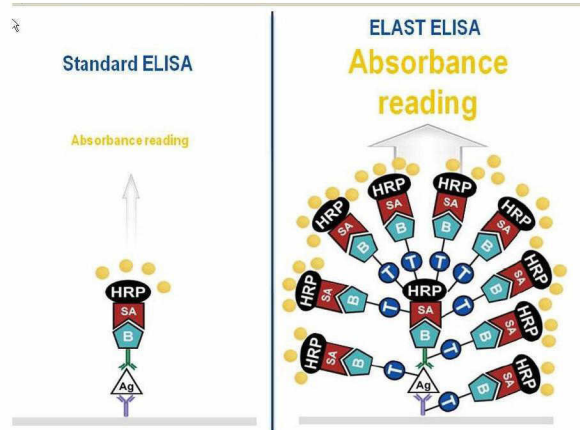


Figure 2. Dissociation enhancement.

Figure 1. Unique fluorescence properties of lanthanides, (a) long fluorescence decay times, (b) large Stokes' shift.

ELAST

- ELISA amplification system
- Signal amplification
- Generally not effective because of background amplification (Contrast PCR)



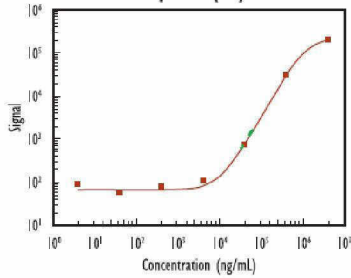
Attributes (good and bad) of selected competing technology

Assay Protocols

- Block MSD MULTI-SPOT plate for 1 hour, wash
- Add 25 μ L of assay diluent solution to each well
- Add 25 μ L of calibrator or sample (undiluted for Panel I; 20-fold diluted for Panel II; 250-fold for PICP assay; and 20-fold for TGF β 1 assay) to each well
- Incubate with shaking for 120 minutes, wash
- Add 25 μ L of labeled antibody solution to each well
- Incubate with shaking for 60 minutes, wash
- Add MSD Read Buffer
- Read plate on MSD Reader

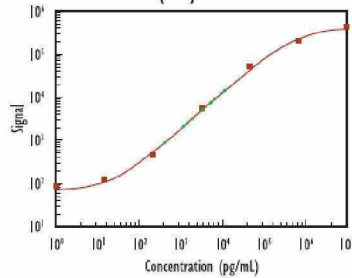
MSD assay performance

Bone Alkaline Phosphatase (ALP) Calibration Curve



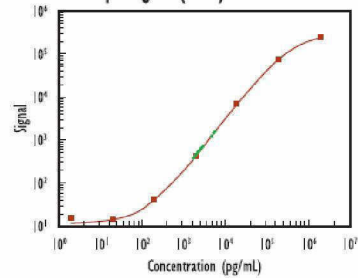
Detection Limit:	8 ng/mL
Dynamic Range:	8-1,000 ng/mL

Sclerostin (SOST) Calibration Curve



Detection Limit:	1 pg/mL
Dynamic Range:	1-10,000 pg/mL

Osteoprotegerin (OPGN) Calibration Curve



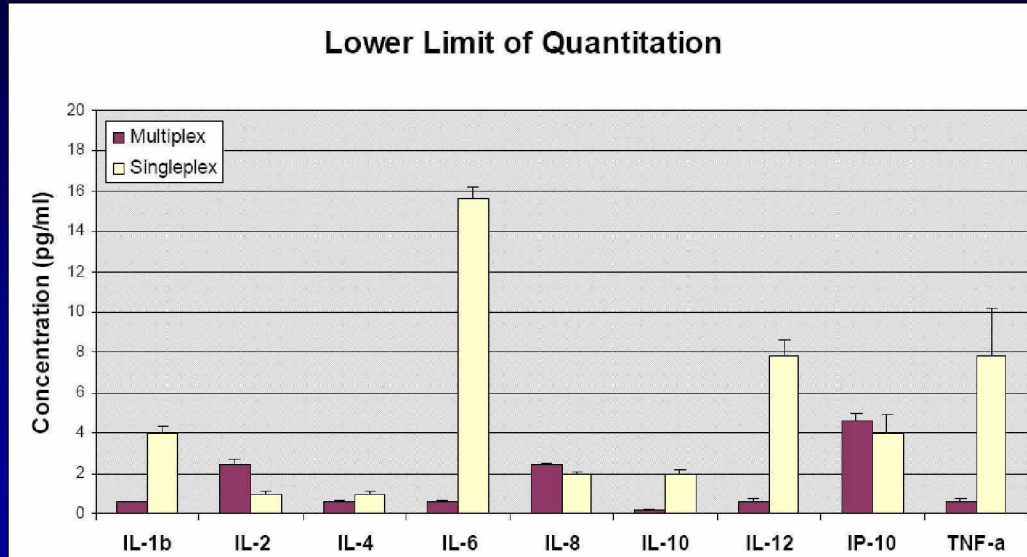
Detection Limit:	6 pg/mL
Dynamic Range:	6-20,000 pg/mL

<http://mesoscale.com/CatalogSystemWeb/WebRoot/literature/applications/pdf/humanBONE.pdf>

Chemical multiplexing

- Coded beads (Luminex, BD, BioRad)
- eTags (Monogram)
- The main problem is complexity and reliability
- In the case of chemical multiplexing background can be a problem

Luminex: Multiplex vs Singleplex



Biosite

Multiplexed (3) IA

Good sensitivity

Blood samples (50 -200 uL)

Precision may not be good enough

Small menu

FDA cleared

Requirements for other types of assay

General chemistry: glucose, lipids

Enzymes: ALT etc.

Electrolytes and blood gasses

Extending the capability of the current system Glucose assay chemistry

Current approach is chemiluminescent

- Compatible with current instrument
- $\text{Glucose} + \text{ATP} \rightarrow \text{Glucose-phosphate} + \text{ADP}$ (Hexokinase)
- $\text{ATP} + \text{D-Luciferin} + \text{O}_2 \rightarrow \text{Oxyluciferin} + \text{AMP} + \text{PPi} + \text{CO}_2 + \text{Light (560nm)}$ (Luciferase)
- Reverse reading assay; measures ATP remaining after ATP is converted to ADP

If this approach works

- Lipid assays and enzyme assays may be feasible

What additional critical analytes do we need/want?

Clinical chemistry

- Glucose (WIP)
- HbA1c (Method proposed; Ab availability is an issue)
- Lipids
 - Cholesterol, HDL-C, LDL-C, Triglycerides
- ALT

White cell markers ??

- CD4, CD8 etc.

Small molecules with no IA chemistry compatible with current system and possible modifications

- Ions (detection means)
- Serotonin (organic chemistry required)

Appendix 2

Selected, existing POC systems for comparison

Biosite: Multiplexed Immunoassay



Cholestech: Multiplexed lipids and very limited other analytes



Abaxis: Multiplexed general chemistry and very limited immunoassay menu



HemoSense: PT/INR (only)



Hemocue: Hemoglobin and glucose



I-Stat: Multiplexed electrolytes and limited immunoassay



Pulse Oximeter

- Measures Hb saturation with O₂
- Non-invasive
- Could easily be added/attached to our instrument





Appendix 3: Imaging Samples

Measurement of sample volume
Determination of sample integrity

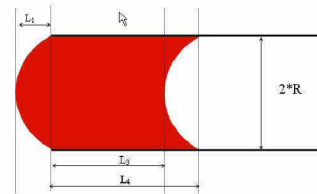
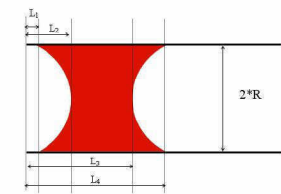
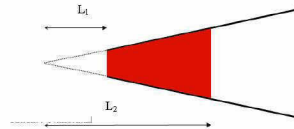
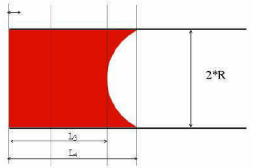
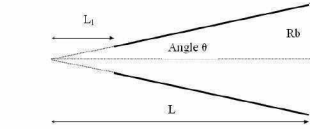
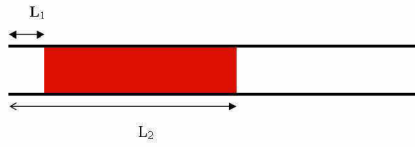
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Imaging enables sample volume measurement

- Verification that the sample has the correct volume
- Verification that sample delivery was good
- Measurement of sample volume
 - Enables result correction for improper volume
- Inexpensive camera will work for this purpose

Some examples



Calculation of volume (example)

Example 4: Geometry of measurement in a conical capillary (often used for tips)

Rb = radius at base of cone

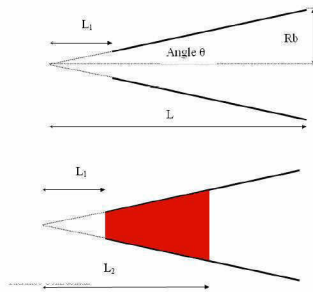
L = length

L1 = distance from (projected) top of the cone to lower sample meniscus

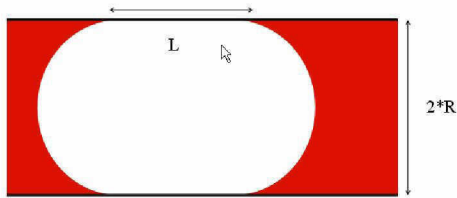
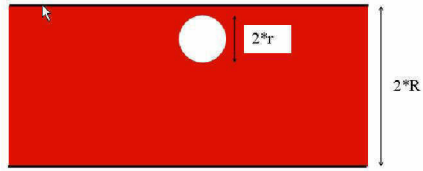
L2 = distance from (projected) top of the cone to lower upper meniscus

Volume introduced = $\rho \cdot (Rb/L)^2 \cdot [(L1)^3 - (L2)^3] / 3$

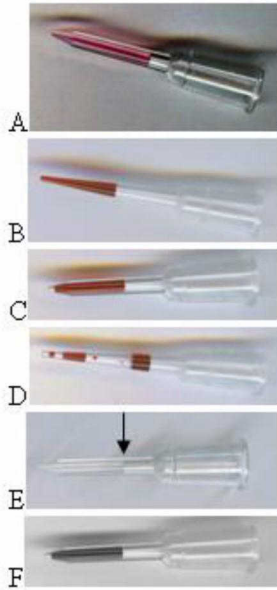
$\tan \theta = Rb/L$



Bubbles



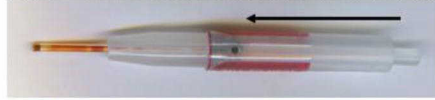
Images



B: Sample transfer device with its capillary filled with sample. The "fill to" location is indicated.



C: Sample transfer device with sample displaced by movement of the plunger



D: Sample transfer device with sample incompletely displaced



Measurement from image of conical capillaries

